#3N03

# U.S. Fish and Wildlife Service Region 3 Contaminants Program

1992 Sediment Quality Report for Mark Twain National Wildlife Refuge Illinois, Iowa and Missouri





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# 1992 Sediment Quality Report for Mark Twain National Wildlife Refuge Illinois, Iowa and Missouri

by Mike Coffey
U.S. Fish and Wildlife Service
Rock Island Field Office
4469 48th Avenue Court
Rock Island, Illinois 61201

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### **Abstract**

Sediment quality was assessed in 1992 by the U.S. Fish and Wildlife Service at six backwater sites along the Upper Mississippi River. Five of the backwater sites were in Mark Twain National Wildlife Refuge. The Sediment Quality Triad approach was attempted to evaluate heavy metal and nutrient contamination. The Sediment Quality Triad methods included analytical chemistry, benthos assessment and toxicity testing using the Microtox system. Heavy metal concentrations were within or slightly elevated above background limits. Nutrient concentrations were slightly to very elevated. The reference site for the study area was at Skunk Slough in Pool 19. Skunk Slough had abundant infaunal macroinvertebrates with above average heavy metal concentrations and low ammonia concentrations. Keithsburg Division in Pool 18 was identified as a backwater with contaminant problems. Keithsburg Division had a poor assemblage of infaunal macroinvertebrates with above average heavy metal concentrations and the highest concentrations of ammonia for the study area. Ammonia toxicity was the suspected cause of some mortality in sediment elutriate toxicity tests completed at Keithsburg Division. The elutriate toxicity tests were intended to validate the Microtox tests. The Microtox® test results were not used because they did not relate to the benthos diversity and chemistry data. GIS analysis indicated that the contaminant problems at the Keithsburg Division backwater were not related to point source pollution such as wastewater discharges into adjacent rivers. The results from this project were used to initiate a multi-year study on nonpoint source pollution at Keithsburg Division.

# **Table of Contents**

Abstract	Ш
	vi
List of Figures	/iii
List of Appendices	İΧ
Acknowledgements	X
Introduction	1
Background	1
Objectives	2
Study Area	3
Study Sites	4
Sampling Locations	6
Methods	7
Sediment Quality	7
Infaunal Macroinvertebrates	9
Toxicity resulting	10
MICIOTOX Test	10
Sediment Liathato 100t 11111111111111111111111111111111	10
water quality	11
Data Analyses	11
Statistical Analysis	11
GIO Allalysis	12
Quality Assurance and Quality Control 11111111111	12
nesults	14
Sediment Quality	14
Sediment roxidio	15
Sediment Chemistry	15
Metals	15
rationts	17
Illiadia Macionivo tebratos	18
Toxicity resting	19
MICIOLOX Test	19
Sediment Liathato 100t	20
water quality	21
Quality Assurance and Quality Control	21

# **Table of Contents - Continued**

Discussion		 						22
Sediment Chemistry		 						22
Metals								22
Nutrients								22
Potential Sources								23
Potential Pathways								23
Infaunal Macroinvertebrates								25
Toxicity Tests								27
Microtox Test								27
Sediment Elutriate Test .		 						28
Sediment Quality Triad								28
Water Quality								28
Conclusions								30
Recommendations								31
Management Recommendations								31
Research Recommendations								32
References								34

# **List of Tables**

<u>Table</u>		<u>Page</u>
Table 1.	Study sites for the Mark Twain National Wildlife Refuge contaminants study, 1992	5
Table 2.	Levee and general land cover information for the study sites, Mark Twain National Wildlife Refuge, 1992	5
Table 3.	Chemical data acquisition plan for the Mark Twain National Wildlife Refuge contaminants study, 1992	8
Table 4.	Water quality parameters and instruments used for the Mark Twain National Wildlife Refuge contaminants study, 1992	11
Table 5.	Average sediment texture data for the study sites, Mark Twain National Wildlife Refuge, 1992	14
Table 6.	Mean, minimum and maximum arsenic and heavy metal concentrations for the study area, Mark Twain National Wildlife Refuge, 1992	15
Table 7.	Average arsenic and heavy metal concentrations for the study area with comparison data, Mark Twain National Wildlife Refuge, 1992	16

# **List of Tables - Continued**

<u>Table</u>		Page
Table 8.	Correlation of arsenic and heavy metal with total organic carbon content and percent clay for the study area, Mark Twain National Wildlife Refuge, 1992	16
Table 9.	Average sediment arsenic and heavy metal concentrations for the study sites, Mark Twain National Wildlife Refuge, 1992	17
Table 10.	Average sediment ammonia and phosphate concentrations for the study area with comparison data, Mark Twain National Wildlife Refuge, 1992	18
Table 11.	Average number of infaunal macroinvertebrates per grab sample for the study sites, Mark Twain National Wildlife Refuge, 1992	19
Table 12.	Average Microtox results for the study sites, Mark Twain National Wildlife Refuge, 1992	20
Table 13.	Results for the sediment elutriate toxicity tests for the study area, Mark Twain National Wildlife Refuge, 1992	20
Table 14.	The ranges of water quality data for the study sites, Mark Twain National Wildlife Refuge, 1992 .	21

# List of Figures

# <u>Figure</u>

Figure 1.	Location of Mark Twain National Wildlife Refuge	2
Figure 2.	Study area and sites for the Mark Twain National Wildlife Refuge contaminants study, 1992	3
Figure 3.	Infaunal macroinvertebrate illustrations	
Figure 4.	View of land cover and use at Keithsburg Division	
Figure 5	Example of changes in dissolved oxygen levels in a productive backwater lake along the Upper Mississippi River	29

# List of Appendices

# **Appendix**

Α	Study Site Maps	<b>A</b> -1
B.	Contaminant and Texture Data Records	
C.	Macroinvertebrate Data Sheets	
D.	Microtox <sup>®</sup> test Reports	
E.	Sediment Elutriate Toxicity Test Reports	
F.	Water Quality Data Records	
G.	Quality Assurance Quality Control Test Results	

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### Introduction

This document reports and interprets sediment quality information for selected Upper Mississippi River backwater areas in Mark Twain National Wildlife Refuge (MTNWR) (Figure 1). The sediment quality information includes analytical chemistry, benthic macroinvertebrate diversity and toxicity test results. Point source and non-point source pollution data were cataloged using Geographical Information System (GIS) methods to help identify potential pathways for contaminants. The project was conducted by biologists from the U.S. Fish and Wildlife Service's Rock Island Field Office (Rock Island, IL) as part of the contaminants program for the Service's Division of Refuges and Wildlife.

### **Background**

In 1989, staff from the Rock Island Field Office surveyed sediment chemistry along the Illinois River and Mississippi River in Illinois, Iowa and Missouri (Young 1991). The 1989 data show that organic pollutants such as polychlorinated biphenyls (PCB's), petroleum products, chlordane and similar pesticide compounds were not detected at MTNWR units along the UMR except for Keithsburg Division (Young 1991). Several hydrocarbon compounds (polycyclic aromatic hydrocarbons - PAHs) were detected at low concentrations at Keithsburg Division (Young 1991). The PAHs at Keithsburg Division are believed to be those produced in natural systems by aquatic plants.

The results of the 1989 study indicated some sites in MTNWR had poor sediment quality conditions based on the mortality of test organisms used in toxicity tests (Young 1991). In 1992, several sites in MTNWR with suspected poor sediment quality were re-evaluated in greater detail to more clearly identify the ecological impacts from contaminants.

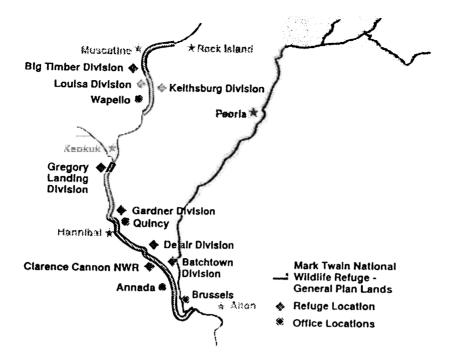


Figure 1. Location of Mark Twain National Wildlife Refuge, Illinois, Iowa and Missouri.

## **Objectives**

The objectives for the re-evaluation of sediment quality are listed below.

- 1. Measure heavy metal and ammonia concentrations in surficial sediments in selected backwater areas.
- 2. Examine the substrate for infaunal (substrate dwelling) macroinvertebrate species that are indicators of good sediment quality.
- 3. Test the toxicity of the sediments with the Microtox system and validate the Microtox tests with elutriate tests.
- 4. Initiate a GIS database with sediment quality and related environmental information.

## **Study Sites**

The refuge sites re-evaluated in 1992 were Batchtown Division (Pool 25), Clarence Cannon Refuge (Pool 25), Gardner (Pool 21), Keithsburg (Pool 18) and Big Timber (Pool 17) Divisions. A non-refuge study site was established in Skunk Slough (Pool 19) to serve as a reference sampling area. The locations of the study sites are outlined in Figure 2 and related geographic information is in Table 1. Study site maps with the positions of the sampling locations are illustrated on site maps in Appendix A.

Skunk Slough was used as a reference sampling area because it supported diverse populations of benthic macroinvertebrates including pollution sensitive species such as burrowing mayflies (*Hexagenia* species) and fingernail clams (family Spaeriidae). The presence of abundant pollution sensitive organisms served as an indication of good sediment quality for this study.

All of the study sites except Clarence Cannon Refuge are forested backwater complexes with old sloughs, lakes and wetlands. Clarence Cannon Refuge is a network of wetlands within a green tree reservoir.

The study sites are directly adjacent to the Upper Mississippi River on one side and upland cover on the other sides. Various levels of protection from floods exist at the different study sites. Some of the sites are hydraulically connected with the Mississippi River via open water channels and others are separated from the river by levees. Some of the sites receive upland run-off and others are protected from run-off by upland levees or forest zones. Refer to Table 2 for an outline of the land features present at each study site.

Table 1. Location information for the study sites, Mark Twain National Wildlife Refuge contaminants study, 1992.

Study Site	Pool	River Mile	County	State
Big Timber Division	17	443	Louisa	lowa
Keithsburg Division	18	428	Mercer	
Skunk Slough	19	396.4	Lee	lowa
Gardner Division	20	332.6	Adams	
Clarence Cannon Refuge	25	260.5	Pike	Missouri
Batchtown Division	25	246	Calhoun	Illinois

Table 2. Levee<sup>1</sup> and general land cover information for the study sites, Mark Twain National Wildlife Refuge, 1992.

Study Site and Acres	River Levee	Upland Levee	Adjacent Land Cover
Big Timber Division (3,375 acres)	No	Yes	Agricultural
Keithsburg Division (1,400 acres)	Yes	No	Agricultural
Skunk Slough (1,500 acres)	No	No	Forest
Gardner Division (6,000 acres)	No	No	Forest
Clarence Cannon Refuge (3,750 acres)	Yes	Yes	Agricultural
Batchtown Division (2,250 acres)	No	No	Agricultural

Backwater is not hydraulically connected to rivers and streams during normal stage

## **Sampling Locations**

We targeted locations in each study site where fine grain sediments are likely deposited. These optimal sampling locations were established at tributary inlets and non-vegetated basins within each site. Several optimal sampling locations were assessed at each study site depending on the size of the backwater and accessibility.

This strategy should identify maximum sediment contaminant concentrations for each site because inorganic contaminants are associated to a greater degree with fine grain sediments. This is because contaminants that are released into surface water generally bind more readily to silt, clay and organic matter. Waterborne contaminated sediments are transported as suspended solids in rivers and streams and are deposited in slack water areas where they settle out of the water column under low flow conditions.







### Methods

The Sediment Quality Triad approach (Long and Chapman 1985; Chapman 1986) was attempted to identify sites with contaminant problems and show if there were ecological impacts from pollution. The Triad approach used three assessment methods. The assessment methods included sediment chemistry, evaluation of infaunal benthos diversity and sediment toxicity testing. The relationships of the results from the assessment methods were used to characterize ecological impacts. Adverse impacts may include low benthos diversity, mortality in toxicity tests and elevated chemical concentrations.

## **Sediment Quality**

Sediments were collected with a standard Ekman dredge with a pole handle. Two sediment grab samples at ten feet apart were taken at each location. The dredge was inserted, closed and raised for inspection. The contents were emptied into a stainless steel bowl if the dredge was at least three guarters full. This included between 13 and 19 centimeters of the sediment column. If it was not a complete grab, another grab was attempted approximately six feet in any direction from the last attempt. The material was gently mixed with a stainless steel spoon and portions were scooped into chemically clean containers for analyses. The samples were chilled in a cooler with blue ice or wet ice. The samples were mailed or transported to the office for storage in a refrigerator or standard chest freezer depending on the type of analyses to be performed. The samples were forwarded to contract laboratories for analysis according to the Chemical Data Acquisition Plan (Table 3). Both grab samples from each location were analyzed for heavy metals and texture data. The first of the two grabs was also analyzed for ammonia. Seventeen archived samples were later analyzed for phosphates.

Table 3. Chemical data acquisition plan for the Mark Twain National Wildlife Refuge contaminants study, 1992.

Analyte	Analytical Method	Quantitation
	Method	Limit
Arsenic	Graphite furnace AA1	$0.5 \ \mu g/g^2$
Cadmium	Graphite furnace AA	0.2 μg/g
Chromium	ICP <sup>3</sup>	1.0 <i>μ</i> g/g
Copper	ICP	1.0 <i>μ</i> g/g
Lead	Graphite furnace AA	5.0 <i>μ</i> g/g
Mercury	Cold vapor AA	0.1 μg/g
Nickel	ICP	5.0 <i>μ</i> g/g
Selenium	Graphite furnace AA	1.0 <i>μ</i> g/g
Zinc	ICP	5.0 <i>µ</i> g/g
Total organic carbon	Coulometrically	
Grain size distribution	Sieve weight	
Ammonia-nitrogen	Phenate Method	0.1 <i>μ</i> g/g
Phosphate-phosphorus	Automated Ascorbic Acid Method	0.1 <i>μ</i> g/g

**Atomic Absorption Spectroscopy** 

<sup>&</sup>lt;sup>2</sup> Micrograms per Gram

<sup>&</sup>lt;sup>3</sup> Inductively Coupled Plasma Emission Spectroscopy

## Infaunal Macroinvertebrates

Sediment grab samples were collected as described above. Care was taken not to dredge the same spot from which the sediment chemistry samples were collected. The contents from the grab were emptied into plastic bags and labeled for storage. In the field, samples were maintained chilled in a cooler with blue ice or wet ice. The samples were mailed or transported to the office laboratory for storage in a standard chest freezer.

A notation was made on the field data sheet if burrowing mayflies or fingernail clams were observed during field handling (eg. stirring or spooning sediment into sample containers) of the sediment samples. The organisms were easy to detect if they were present above low numbers because of their size and movement. The fingernail clams were not active directly after collection, but were large enough by May for easy detection. Mayflies were easy to detect because they constantly moved about and fanned their tail and gill structures.

At least two frozen samples from each site were randomly selected and processed for macroinvertebrates. The contents of each plastic bag were completely emptied into a benthos bucket (Wildco Company) with a mesh size of 0.595 millimeters. The sediment was washed from the debris and organisms with tap water through a garden hose. The material trapped by the sieve was backwashed into a white enamel pan.

Fingernail clams with articulated valves, burrowing mayfly nymphs and other recognizable taxonomic groups present in the material were enumerated and the sample discarded.

## **Toxicity Testing**

Sediment grab samples were collected as described above. Care was taken not to dredge the same spots from the sediment chemistry and macroinvertebrate samples were collected. The samples were maintained in a cooler with blue ice or wet ice. The samples were forwarded to the office laboratory for storage in a standard chest freezer for Microtox testing and in a refrigerator for the sediment elutriate toxicity test.

The Microtox<sup>®</sup> solid phase sediment assay test was performed on sediments from all of the sites to rank the relative toxicity between the sites. Elutriate toxicity tests were performed on sediments from one site (Keithsburg Division) to help interpret the results of the Microtox<sup>®</sup> test.

## Microtox<sup>®</sup> Test

Microtox assays were performed in duplicate on at least two samples randomly selected from each site. The assays were completed at the Rock Island Field Office laboratory. The Microbics version 6.3 detailed solid-phase test was used (Microbics, 1989 et. seq.). Room temperature, color and turbidity were monitored.

### **Sediment Elutriate Test**

The University of Iowa Hygienic Laboratory performed the elutriate toxicity tests. Sediments from Keithsburg Division were tested because metal and ammonia concentrations were elevated for the study area. The toxicity tests were duplicate 96- hour static acute bioassays. A 200 milliliter volume of sediment was thoroughly mixed with purified laboratory water at a ratio of 1:4. The elutriate was allowed to settle for about 60 hours before the introduction of larval fathead minnows (*Pimephales promelas*).

The bioassay water was aerated and the fish were fed brine shrimp on days three and five. The bioassay water was monitored for temperature, pH, dissolved oxygen, ammonia. Unionized ammonia concentrations were calculated for each bioassay vessel.

## **Water Quality**

Selected water quality measurements were taken at approximately 0.3 meters (one foot) below the surface and approximately 0.3 meters above the substrate if greater than three meters deep. Table 4 lists the water quality parameters and instruments used.

Table 4. Water quality parameters and instruments used for the Mark Twain National Wildlife Refuge contaminants study, 1992.

Parameter	Instrument Units	Instrument	Precision
Depth	meters	Marked pole	
Temperature	degrees celsius	YSI Meter Model 51	
Dissolved oxygen	milligrams/liter	YSI Meter Model 51B	25% of scale
Conductivity	$\mu$ S/centimeter	YSI Meter Model 33	2% of scale

## **Data Analyses**

## Statistical Analysis

Descriptive statistics of the contaminants data were calculated using Excel version 4.0 (Microsoft Corporation). Descriptive statistics included arithmetic mean, standard deviation, minimum and maximum values. Correlation coefficients (r) for the chemistry and texture data were calculated using Excel.

The results of the Microtox<sup>®</sup> assays were analyzed using Microtox<sup>®</sup> software version 6.3 (Microbics Corporation) to determine the EC<sub>50</sub> which is the concentration at which effective response by the bacteria was reduced by 50%.

## **GIS** Analysis

The spatial relationships of the contaminants and environmental data were analyzed using Environmental Planning and Programming Language version 7.0 (EPPL7) (State of Minnesota, Land Management Information Center, St. Paul, MN).

The environmental data included land cover, hydrography and industrial wastewater discharge points for MTNWR. The land cover was available from UMR floodplain satellite imagery attributed by the National Biological Survey's Environmental Management Technical Center (Onalaska, WI). The wastewater discharge locations were gathered from state inventories of National Point Discharge Elimination System permits (NPDES).

## **Quality Assurance and Quality Control**

Field data sheets were completed at the time of sampling and included information on water quality, aquatic plant growth, substrate condition, date, time, weather and collector's name.

The position of the sampling locations were plotted on detailed refuge maps. The points were later entered on the GIS base map and Universal Transverse Mecator coordinates (UTM) were obtained for the sampling locations.

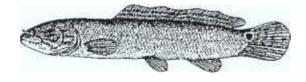
The instrument probes and collection gear were thoroughly rinsed with river water, then acetone (if appropriate for the gear) and de-ionized water between uses.

The dissolved oxygen meter was calibrated by the air calibration technique. The precision of the field equipment are listed in Table 4.

The contract analytical laboratories conducted quality assurance and quality control tests with six randomly selected samples and reference media. The tests included procedural blank samples, replicate tests, testing reference materials and spike recovery analysis.

Control and reference toxicant bioassays were conducted with the elutriate toxicity tests.





### Results

## **Sediment Quality**

#### **Sediment Texture**

The dominant grain sizes at the sampling stations were silt and clay. Sediment total organic carbon content concentrations (TOC) were less than five percent with Keithsburg Division samples in the high part of this range. Average texture data for the study sites are outlined in Table 5.

One sampling location out of the total of 85 had a high percentage of sand (88 percent). This location was in a small bay of Big Timber Division adjacent to upland sandy soils. The contaminant data from this location was excluded from descriptive and correlation analyses because inorganic pollution is generally not associated with substrates of quartz sand (Brannon *et al* 1976). The texture data from each sampling station are listed on copies of the original spreadsheets in Appendix B.

Table 5. Average sediment texture data for study sites, Mark Twain National Wildlife Refuge, 1992.

Study Site	%Organic	%Clay	%Silt	%Sand
Skunk Slough	2.4	45.85	49.07	
Big Timber Division	1.95	33.61	51.69	
Keithsburg Division	3.54	36.09	52.13	11.79
Gardner Division	1.36	17.68	59.18	23.14
Clarence Cannon Refuge	2.52	47.31	45.72	
Batchtown Division	1.93	39.15	53.05	7.38

## **Sediment Chemistry - Metals**

The average sediment concentrations of arsenic and heavy metals for the study area are listed in Table 6. Selenium and mercury were not detected. The mean metal concentrations for the study area except for zinc were within background concentrations for soils and aquatic sediments in Illinois (Table 7). Zinc was 1.5 times greater than average background concentrations for soil and 1.7 times greater for aquatic sediments in Illinois. The correlation of arsenic and heavy metals with grain size distribution and TOC content are provided in Table 8. The best associations were copper with percent clay (r=0.77) and cadmium with TOC (r=0.63). The average concentrations of arsenic and heavy metals varied slightly between the sampling sites with concentrations exceeding background at some sampling locations (Table 9). The chemistry data from each sampling station are listed in Appendix B.

Table 6. Mean, minimum and maximum arsenic and heavy metal concentrations in sediments (micrograms per gram, dry weight, n=83) for the study area, Mark Twain National Wildlife Refuge, 1992.

Analyte	Mean (n=83)	Standard Deviation	Minimum	Maximum
Arsenic	5.32	1.61	0.5	8.18
Cadmium	0.45	0.17	ND¹	0.77
Chromium	18.53	9.02	3.69	50.52
Copper	21.05	6.01	7.70	34.65
Nickel	21.36	8.15	ND	44.30
Lead	21.19	7.07	7.82	51.92
Zinc	83.62	29.36	16.00	145.6

ND = not detected above quantitation limit for this analyte

Table 7. Average arsenic and heavy metal concentrations in sediments (micrograms per gram, dry weight) for the study area with comparison data, Mark Twain National Wildlife Refuge, 1992.

Analyte	Study Area		Soils		Background Sediments		
	Mean	Max	MN¹	U.S. <sup>2</sup>	Great Lakes <sup>3</sup>	IL Streams⁴	
Arsenic	E 22	0.10		•		<b>40</b>	
Cadmium	5.32 0.45	8.18 0.77	0.3		<3	<8 <0.5	
Chromium		****		F.0	- 0E		
	18.53	50.52	43	53	<25	<16	
Copper	21.05	34.65	26	25	<25	<38	
Nickel	21.36	44.30	21	20	<20		
Lead	21.19	51.92	<25	20	<40	<28	
Zinc	83.62	145.6	54	54	<90	<80	

Average Minnesota soil profiles (n = 24) (Baily and Rada 1984)

Average elemental concentrations in surficial materials (Schacklette *et al* 1971)

Average concentrations for non-polluted harbor sediments in the Great Lakes (n = 260, USEPA 1977).

Non-elevated stream sediments in Illinois (n=79, IEPA 1984).

Table 8. Correlation<sup>1</sup> of arsenic and heavy metals with total organic carbon content and percent clay for the study area, Mark Twain National Wildlife Refuge, 1992.

	As²	Cd	Cr	Cu	Ni	Pb	Zn
%TOC³	0.20	0.63	0.15	0.47	0.18	0.26	0.29
%Clay	0.33	0.53	0.28	0.77	0.47	0.50	0.5

The correlation coefficient (r) is a measure of the closeness of the relationship between two variables, r=1 indicates a perfect relationship Arsenic, cadmium, chromium, copper, nickel, lead and zinc, respectively Percent total organic carbon content

Table 9. Average sediment arsenic and heavy metal concentrations (micrograms per gram, dry weight) for the study sites, Mark Twain National Wildlife Refuge, 1992.

Study Site	As <sup>1</sup>	Cd	Cr	Cu	Ni	Pb	Zn
Skunk Slough (n = 6)	6.60	0.43	17.12	28.94	21.28	30.30	92.73
Big Timber Division (n = 16)	5.39	0.42	21.58	21.51	25.05	26.45	108.11
Keithsburg Division (n = 14)	6.10	0.57	20.66	22.52	21.07	21.61	89.53
Gardner Division (n = 8)	4.35	0.38	10.58	11.45	9.87	11.76	45.92
Clarence Cannon Refuge (n = 15)	3.71	0.35	13.05	20.09	17.21	17.87	66.10
Batchtown Division (n=24)	5.83	0.49	21.67	21.71	26.52	20.39	85.08

Arsenic, cadmium, chromium, copper, nickel, lead and zinc, respectively

## **Sediment Chemistry - Nutrients**

The average concentrations of sediment nutrients from the study sites are listed in Table 10. Sediment ammonia concentrations were slightly to very elevated as compared to background sediment chemistry data (Table 10). Keithsburg Division had the highest concentrations in the study area of sediment ammonia and phosphates (average total ammonia-nitrogen = 231.43 micrograms/gram; phosphate = 2000.0 micrograms/gram). Ammonia concentrations had good correlation with TOC (r=0.75). The chemistry data from each sampling station are listed in Appendix B.

One sampling station at the south end of Keithsburg Division had a bulk sediment ammonia concentration of 1400 micrograms per gram, dry weight. The results from this sampling station were excluded from the calculations for the descriptive and graphical statistics because it was so much higher than the ammonia data from the other seven sampling stations at Keithsburg Division.

Table 10. Average sediment ammonia and phosphate concentrations (micrograms per gram, dry weight) for the study area with comparison data, Mark Twain National Wildlife Refuge, 1992.

Sampling location	Ammonia-nitrogen	Phosphate		
Skunk Slough	81.66 (n = 3)	1100.0 (n = 1)		
Big Timber Division	103.00 (n = 7)	795.0 (n = 4)		
Keithsburg Division	231.43 (n = 7)	2000.0 (n = 2)		
Gardner Division	samples lost	samples lost		
Clarence Cannon Refuge	102.40 (n = 5)	742.5 (n = 4)		
Batchtown Division	83.30 (n = 6)	846.7 (n = 6)		
Non-polluted <sup>1</sup>	75.00 (n = 260)	1369.0 (n = 260)		

The means were used to classify non-polluted aquatic sediments from Great Lakes harbors between 1974 and 1975 (USEPA 1977).

#### Infaunal Macroinvertebrates

Midge larvae (families Chironomidae and Heleidae) and aquatic worms (class Oligochaeta) were observed throughout most of the study area.

In addition to midges and aquatic worms, abundant burrowing mayfly nymphs together with fingernail clams were collected at the reference site (Skunk Slough) and abundant burrowing mayflies were collected at Gardner Division. Infaunal macroinvertebrates were poorly represented in the sediment samples from Keithsburg Division.

The number and types of organisms collected at the study sites are outlined in Table 11. The macroinvertebrate data from each sampling station are listed in Appendix C.

Table 11. Average number and the taxa of infaunal macroinvertebrates per grab sample for locations at the study sites at Mark Twain National Wildlife Refuge, 1992.

Study Site	Midge larvae Chironomidae	Fingernail Clams Spaeriidae	Burrowing Mayflies Hexagenia
Skunk Slough (n = 4)	2.25	3.25	3.5
Big Timber Division (n = 10)	3.7	0	0.1
Keithsburg Division (n = 5)	0.2	0	.01.
Gardner Division (n = 2)	8.5	0.5	3
Batchtown Division (n = 3)	8.7	0.3	0

## **Toxicity Testing**

### Microtox<sup>®</sup> Test

The average effective concentrations ( $EC_{50}$ s) for the study sites are listed in Table 12. The  $EC_{50}$  value is the concentration at which 50 percent of the organisms exhibited a response for each of the Microtox<sup>e</sup> assays. The lower  $EC_{50}$  values are considered to represent a more contaminated media, for it takes less material for a response. There was a good correlation with test results and sand grain size (r = 0.97).

The results suggest that the sediments from some of the sampling locations produced a response by the test organisms and the relative magnitude of the response as measured with this organism varied greatly. Mean EC<sub>50</sub> value from Skunk Slough was measured as the most potent compared to the other study sites. Copies of the Microtox<sup>e</sup> program printouts are in Appendix D.

Table 12. Average Microtox<sup>®</sup> effective concentrations (EC<sub>50</sub>), micrograms per gram) for the study sites at Mark Twain National Wildlife Refuge, 1992.

Study Site	Mean EC <sub>50</sub>	SD	Minimum	Maximum
Skunk Slough	664.84 (n = 4)	213.28	334.85	2659.3
Big Timber Division	3128.72 (n = 4)	2359.88	1417.03	6581.97
Keithsburg Division	1454.83 (n = 4)	358.33	1011.78	1881.13
Gardner Division	6574.05 (n = 4)	85 <b>9</b> 3.05	544.98	16,236.97
Clarence Cannon Refuge	2267.16 (n = 7)	975.58	842.85	3441.76
Batchtown Division	1228.42 (n = 8)	1234.82	343.2	3788.81

### **Sediment Elutriate Test**

There was zero percent mortality in July and up to 20 percent in September of the larval fish used in the elutriate tests at Keithsburg Division (Table 13). Bioassay water quality in the test vessels were normal except for un-ionized ammonia (see data in Table 13). The concentrations of un-ionized ammonia in four out of the six tests at the 48 hour time exceeded lethal concentrations for fathead minnows (0.70 milligrams per litermg/L, 48 hours  $LC_{50}$ ) (USEPA 1985). Copies of the laboratory results for each test are in Appendix E.

Table 13. Results for the sediment elutriate toxicity tests for the study area, Mark Twain National Wildlife Refuge, 1992.

Test	Мо	rtality <sup>1</sup>	Un-ioni	(milligrams per liter)	
	Sample	Control	Initial	48 Hours	96 Hours
7-14-92 A	0/20	0/20	0.12	0.43	0.16
7-14-92 B	0/20	1/20	0.12	0.40	0.20
9-17-92 A	4/20	0/20	0.18	0.92	0.34
9-17-92 B	1/20	0/20.	0.19	1.2	0.34
9-17-92 C	4/20	0/20	0.27	1.0	0.33
9-17-92 D	4/20	0/20	0.22	1.0	0.35

<sup>&</sup>lt;sup>1</sup> Mortality = number dead / number tested

## **Water Quality**

Water quality parameters measured for this project were similar throughout the study area except for dissolved oxygen (DO). Dissolved oxygen was good during most sampling trips except for later trips to Keithsburg Division. Dissolved oxygen levels were low at some locations during July and September at Keithsburg Division. The ranges for the water quality data are summarized in Table 14. The water quality data from each of the sampling locations are listed in Appendix F.

Table 14. The ranges of water quality data for the study sites at Mark Twain National Wildlife Refuge, 1992.

Study Site	Month	Depth (Meters)	Temperature (Celsius)	Conductivity (µS/cm)	Dissolved Oxygen (Milligrams/liter)	
				***	Тор	Bottom
Skunk Slough	Jun	1.0-2.5	26.0-27.0	440-470	6.2-10.0	
Big Timber Division	May/Jun	0.5-5.5	23.0-25.0	310-400	7.5-14.0	5.8-6.2
Keithsburg Division	May	2.75-5.5	20.2-22.5	310-340	5.2-15.0	3.4-12.0
	Jul	3.0-4.0	25.5-26.0	280-315	4.1-4.4	0.5-3.6
	Sep	2.5	19.0-20.0	350-470	2.0-5.7	0.6-4.0
Gardner Division	May	1.5-1.75	19.5-20.5	310-455	8.1-9.2	
Clarence Cannon Refuge	May	0-11.0	17.0-19.0	190-430	7.5-12.0	
Batchtown Division	May	1.5-4.5	20.5-23.0	285-335	5.4-15.0	11-15.0

## **Quality Assurance and Quality Control**

Quality assurance performance results for the sediment analyses conducted by U.S. Fish and Wildlife Service contract laboratories were accepted by the Service's quality control office at the Patuxant Analytical Control Facility (PACF). There were slight differences in the replicate test results. The results of the internal laboratory replicate, spike and procedural blank analyses are provided in Appendix G. The contract laboratories' analytical results and original field data sheets are held at the Rock Island Field Office.

## **Discussion**

## **Sediment Quality**

#### Metals

Mean concentrations of arsenic and heavy metal contaminants were not at levels of concern. Skunk Slough was a site with above average heavy metal concentrations for the study area and both pollution sensitive species of infaunal macroinvertebrates were present in abundance. Skunk Slough was also the site demonstrating an effective response with the Microtox® toxicity test. This relationship provides evidence that these concentrations of arsenic and heavy metals may not be limiting benthic macroinvertebrate production in Refuge backwaters and that the Microtox® results are inconclusive.

#### **Nutrients**

Sediment ammonia concentrations between the low and high ends of the range for the study area may be limiting aquatic life in MTNWR backwaters. Ammonia concentrations in Skunk Slough were near the low end of the range for the study area and that backwater supported abundant pollution-sensitive macroinvertebrates. Sediment ammonia concentrations at Keithsburg Division were at the high end of the range for the study area and infaunal macroinvertebrates were poorly represented there. Sediment ammonia concentrations at Keithsburg Division reached toxic levels for fish as indicated by the elutriate toxicity tests.

Ammonia that is found in the substrate is mostly bound to surrounding fine grain particles in the form of ammonium  $(NH_4^+)$ . A portion of the ammonia is in solution as un-ionized ammonia  $(NH_3)$ . The un-ionized ammonia accumulates within the sediment

pore water and is released at the substrate-water interface. Unionized ammonia is toxic to aquatic organisms at very low concentrations (USEPA 1986). Un-ionized ammonia is generally less toxic to invertebrates (lethal concentration for midge larvae, *Chironomus* sp., is  $LC_{50}=6.6$  mg/L for 96 hours] as compared to fish (lethal concentration for bluegill, *Lepomis* sp., is  $LC_{50}=1.75$  mg/L for 96 hours] (USEPA 1976 and 1985). The fraction of unionized ammonia is calculated using temperature and pH.

Phosphates in sediments are not typically toxic to aquatic animals (USEPA 1986). The significance of high phosphates is the effect of causing nuisance aquatic plant blooms and the resulting build-up of organic matter (USEPA 1986). Aquatic plant blooms can limit dissolved oxygen from excessive plant respiration relative to oxygen production through photosynthesis and by the microbial decay of dead plant tissue.

#### **Potential Sources - Nutrients**

Keithsburg Division had higher sediment nutrient concentrations and higher organic content compared to the other study sites in MTNWR (Table 10). There are likely three sources for nutrients at Keithsburg Division. The first source is ammonia produced in lake sediments from decaying organic matter. Agricultural fertilizer run-off is an important source for nitrogen and phosphorus compounds. The third source is from municipal and industrial wastewater point discharges.

Organic matter that is deposited in the substrate of aquatic and wetland systems is decomposed by bacteria which produce ammonia as a by-product. The ammonia is converted first to nitrites then to nitrates by bacteria (Rand and Petrocelli 1985). The amount of ammonia production depends on the volume and quality of the organic matter and overlying water quality conditions (Rand and Petrocelli 1985). The conversion to nitrate

can consume significant amounts of dissolved oxygen from the water column, especially during the winter after a season of ammonia production (Knowles and Lean 1987).

## **Potential Pathways - Nutrients**

Analysis of the spatial data for Keithsburg Division demonstrated the use of GIS for contaminants investigations and helped identify contaminant pathways. Figure 3 is GIS map product to display the land cover around Keithsburg Division. Overlay data for the spatial analysis included wastewater discharge points and tributary rivers and streams.

There was only one wastewater discharge point near Keithsburg Division. It is the discharge into the Mississippi River for the municipal sewage treatment plant for New Boston, IL. This source is separated from the Keithsburg Division backwater by the river levee. Surface water from the Mississippi River enters the backwater only during flood stages. The 1993 flood on the Mississippi River overtopped the river levee for much of the summer and created a scour route to Pope Creek from Keithsburg Division. Solids and contaminants transported in the flood water may have changed sediment quality at Keithsburg Division.

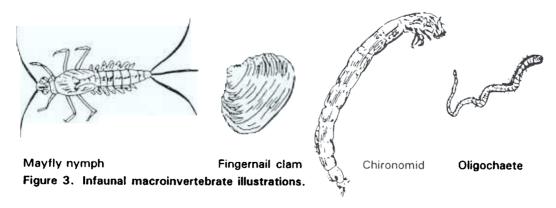
There are no upland levees at Keithsburg Division. The upland cover adjoining Keithsburg Division is agricultural. Surface runoff from adjacent agricultural fields may carry contaminants to the site. The primary sources of surface water for Keithsburg Division are the Edwards River to the north, small un-named tributaries at the north boundary and Pope Creek to the south. The Edwards River and Pope Creek are slightly to moderately impaired from suspended solids, nutrients and habitat modifications (IEPA 1994). Inputs from the Edwards River and Pope Creek may lead to poor water quality at Keithsburg.

## Infaunal Macroinvertebrates

In-lake production of ammonia from organic enrichment likely limits benthic communities at Keithsburg Division backwaters. Infaunal macroinvertebrates were sparse at Keithsburg Division in May of 1992. Poor sediment quality and eutrophication were evidenced at Keithsburg Division by the abundant aquatic plant growth, low dissolved oxygen, high organic content and sediment ammonia toxicity.

Ammonia toxicity has been blamed for the decline of fingernail clams in the Illinois River and the Mississippi River. Highly eutrophic backwater lakes should shift from having diverse macroinvertebrate populations to populations of only a few pollution-tolerant organisms. Sustaining macroinvertebrate production in Mississippi River backwaters is important because they represent critically important food items for migratory birds and fish. Fingernail clams may be locally important to waterfowl.

The absence of fingernail clams and burrowing mayflies in backwater sites may be explained by reasons other than sediment pollution. Macroinvertebrate communities may have a random distribution and their absence does not necessarily indicate poor quality habitat. Backwaters with high sediment organic content may be unsuitable for burrowing mayfly nymphs However, fingernail clams are typically found in shallow, rich backwaters along the Upper Mississippi River.



## **Toxicity Testing**

## Microtox<sup>®</sup> Test

The Microtox<sup>®</sup> assay was used to test for toxicity of backwater sediments. This system is relatively new, inexpensive to run and easy to use. The Microtox<sup>®</sup> system measures luminescence of the marine bacterium *Photobacterium phosphoreum*. Inhibition of this luminescence is considered a toxic response. Results of the Microtox<sup>®</sup> assay compare to other standard freshwater toxicity tests using fish and cladocerans (Kaiser and Palabrica 1991). Natural toxins like sulfur in aquatic sediments are also lethal to this species of bacteria (Jacobs *et al* 1992). *P. phosphorem* are not sensitive to ammonia (Ankley *et al* 1990). The heavy metals and free sulfur from sulfide compounds commonly found in the deeper and anoxic part of the sediment column may have contributed to the effect measured using *P. phosphoreum* for this study.

The results also demonstrated that fine grain sediments caused a greater response than coarse grain sediments. Coarse grain sediments like sand and large silt sizes do not have the same capacity to bind contaminants and generally do not hold high concentrations of inorganic pollutants (USEPA 1976). Fine grain sediments increased the turbidity of the sample and may have affected the measurement of the light output.

The Microtox<sup>®</sup> test did not compare well to the sediment elutriate test because of the presence of ammonia. The Microtox<sup>®</sup> organism is not sensitive to ammonia and fish species are very sensitive to ammonia. Overall, the Microtox<sup>®</sup> test did not adequately analyze the toxicity of backwater sediments because of the role of ammonia toxicity and turbidity interferences.

### Sediment Elutriate Test

Ammonia concentrations were high in the bioassay water for the elutriate toxicity tests at Keithsburg Division. Sediment bound ammonia is easily released in the water column when mixed (Brannon et al 1978). Heavy metals bound to sediments tend not to be released at full concentrations into the water column under neutral and toxic conditions when mixed (Brannon et al 1978).

## **Sediment Quality Triad**

The data from the Microtox tests may not be used for the Sediment Quality Triad analysis because the Triad is an effects-based approach incorporating measures of chemistry, benthos diversity and toxicity (Chapman *et al* 1992). The Triad analysis depends on satisfactory toxicity test results to rank degradation between study sites (Chapman *et al* 1991).

The elutriate toxicity tests for Keithsburg Division were intended to validate the Microtox tests. The elutriate tests were used with benthos data in the Triad approach for this site. There was evidence that the toxicity test results were related to ammonia contamination. This may explain the poor benthos diversity observed at Keithsburg Division.

## **Water Quality**

Water quality was measured only during sediment sampling trips to provide information to help interpret of the sediment chemistry data. The levels of dissolved oxygen at Keithsburg Division were at times below criteria intended to protect native fish communities (criteria = 5 mg DO/L).

Dissolved oxygen levels may vary greatly during the day and throughout the season in backwater lakes with high primary production (Owens and Crumpton 1993). Refer to Figure 5 for an illustration of dissolved oxygen cycles in a productive backwater lake of the Upper Mississippi River. The lowest oxygen levels typically occur near daybreak due to overnight respiration of phytoplankton and other aquatic plants. Supersaturated oxygen conditions typically occur in the afternoon as a result of photosynthesis.

The importance of the low dissolved oxygen observations is that it supports the expected affects of oxygen demand by high plant respiration and organic matter decay related to nutrient enrichment found at Keithsburg Division.

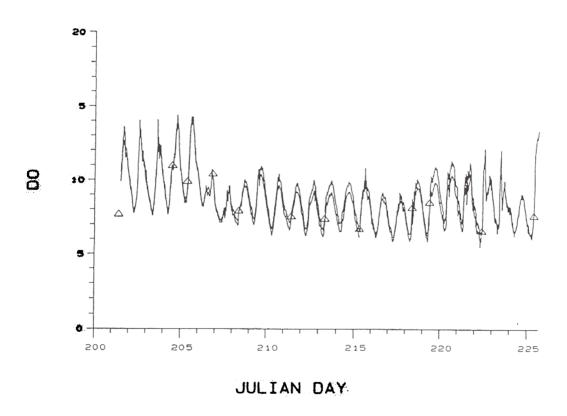


Figure 5. Example of dissolved oxygen levels (DO in parts per million) at a productive backwater lake along the Upper Mississippi River (taken from Owens and Crumpton 1993).

## Conclusions

Prior to the greater than one hundred year flood in 1993, MTNWR backwater sites along the UMR were not grossly polluted with organic pollutants and heavy metals (Young 1991 and this study).

Skunk Slough in Pool 19 was used as a reference site for this study because it supported abundant pollution-sensitive macroinvertebrate species. Skunk Slough had above average concentrations of sediment heavy metals and low sediment ammonia concentrations for the study area.

In contrast to Skunk Slough, ammonia toxicity, nutrient and organic enrichment were found in the backwater habitats of Keithsburg Division. Keithsburg Division also had above average sediment heavy metal concentrations. This site had the highest ammonia concentrations in the study area. There was some mortality in the elutriate toxicity tests conducted later in the season and infaunal macroinvertebrates were poorly represented at Keithsburg Division. GIS analysis indicated that the sources and pathways for contaminants at Keithsburg Division were related to non-point source pollution versus point source pollution.

The methods used for the Sediment Quality Triad approach provided worthwhile data to help describe contaminant-related problems at Keithsburg Division. However, the Microtox® toxicity tests from throughout the study area were inconclusive. The Microtox® test results were not used with the chemistry and benthos data to help describe sediment quality at the other study sites according the Sediment Quality Triad approach. We found that the Microtox® test results did not relate to benthos diversity and sediment toxicity because of the role of ammonia and turbidity in the test samples.

### Recommendations

## **Management Recommendation**

We recommend that the Division of Environmental Contaminants and Division of Refuges formulate strategies to help manage non-point source pollution in backwater habitats of MTNWR. Non-point source pollution includes increased sedimentation and nutrient loading. Nutrient enrichment promotes ammonia production and leads to cultural eutrophication. There are many nutrient management strategies available to reduce the effects from non-point source pollution. These strategies may be grouped into two categories and include on-refuge and off-refuge activities.

On-refuge strategies include diversion of inputs and management of refuge sections as chemical treatment wetlands. Treatment wetland technology is used for nutrient management at municipal sewage treatment plants. Treatment wetlands can assimilate some nutrients and trap solids.

Off-refuge strategies include increasing riparian buffer zones, reducing run-off potential and instituting integrated pest management programs in the watershed.

Many of these strategies may be incorporated into habitat restoration projects by the Environmental Management Program and Farm Bill programs.

No one pollution management method will remove all types of nutrients and filter chemicals continuously. Integrating strategies specific to each site will help protect migratory birds and fish resources. These strategies should improve water quality, generate diverse aquatic macrophyte communities and optimize production of forage resources.

#### **Research Recommendations**

1 Catalog the sediment quality data into the GIS database for MTNWR. The sediment quality database currently under construction by the National Biological Survey for the UMR may be another appropriate repository for these data.

The 1989 and 1992 studies at MTNWR provide baseline data for management purposes and pre-spill conditions for a large U.S. Fish and Wildlife Service refuge. These data are needed for natural areas located along transportation routes for hazardous materials. The transportation routes in MTNWR include barge traffic on the UMR, gas pipelines at Keithsburg Division and various road and railroad bridges over tributaries leading to MTNWR units. The baseline data are available for natural resource damage assessments in the event of a spill.

2. Repeat biomonitoring at MTNWR every five years to evaluate long-term trends in pollution.

The Sediment Quality Triad with modifications is the recommended approach to assess trends in pollution. Many different types of contaminants are deposited in the substrate and sediment quality data provided a status report on contamination to individual backwater systems. Using the Triad approach provided useful ecological data for resource managers. The biomonitoring data are also needed by three other programs under development by the U.S Fish and Wildlife Service. The programs include long-term biomonitoring of refuges (Biomonitoring Environmental Status and Trends - BEST), the migratory bird strategy for the Upper Mississippi River and ecosystem management plans for the Upper Mississippi River.

The modifications to the Sediment Quality Triad approach include changing to ammonia sensitive toxicity tests (see Dillon and Ross 1990) and designing statistically defensible methods.

3. Identify non-point source pollution problems for MTNWR and develop management strategies.

We used the nutrient pollution data and GIS results from this project as an indication of risk from agricultural chemical pollution such as herbicides and insecticides. A risk assessment funded by the Divisions of Refuges and Ecological Services was initiated at Keithsburg Division in 1993 and continues in 1995. The risk assessment will evaluate all of the pathways to the site and provide specific management alternatives to reduce the adverse effects from non-point source pollution.

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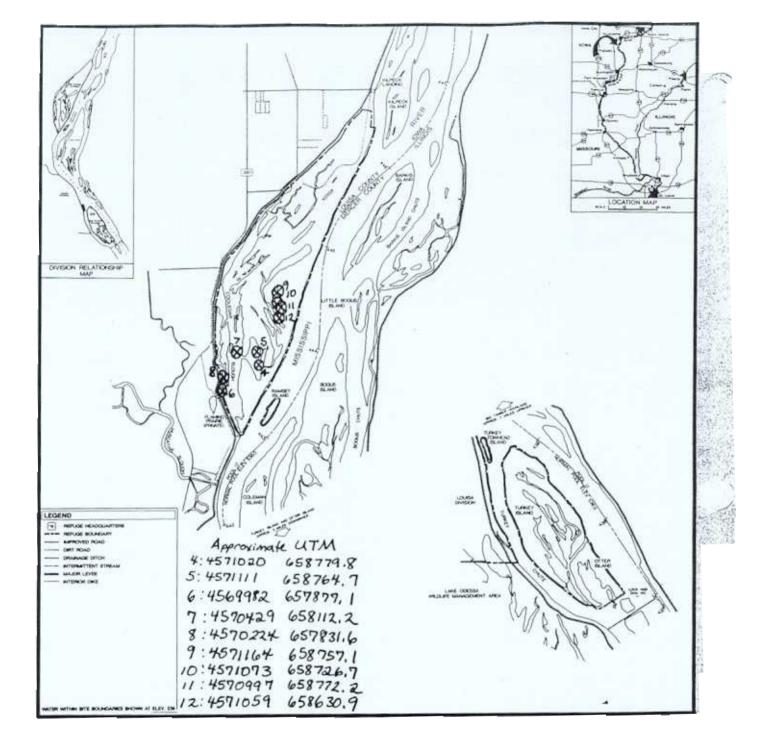
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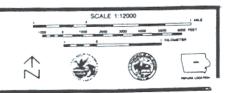
# Appendix A Study Site Maps

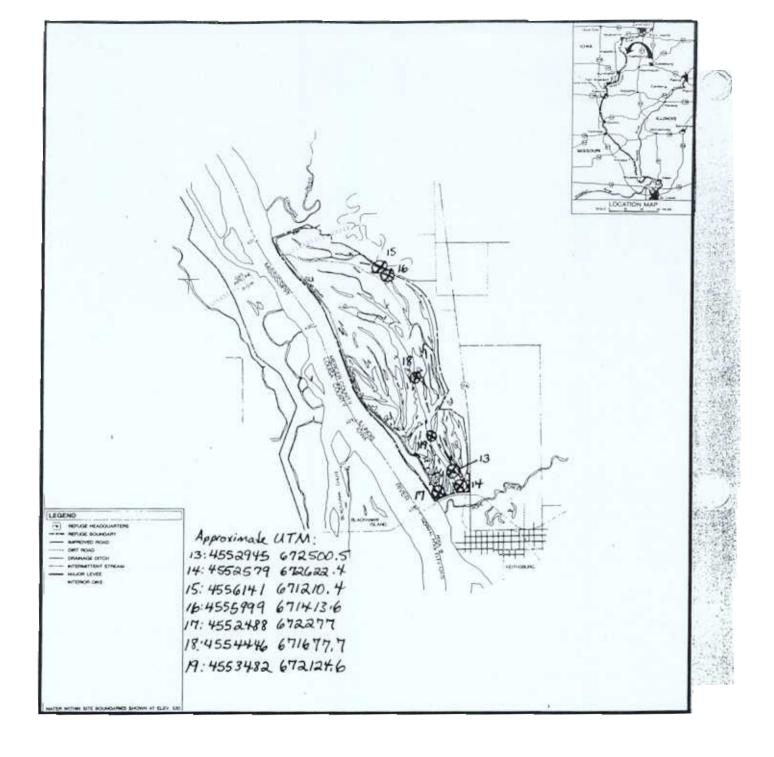




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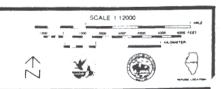
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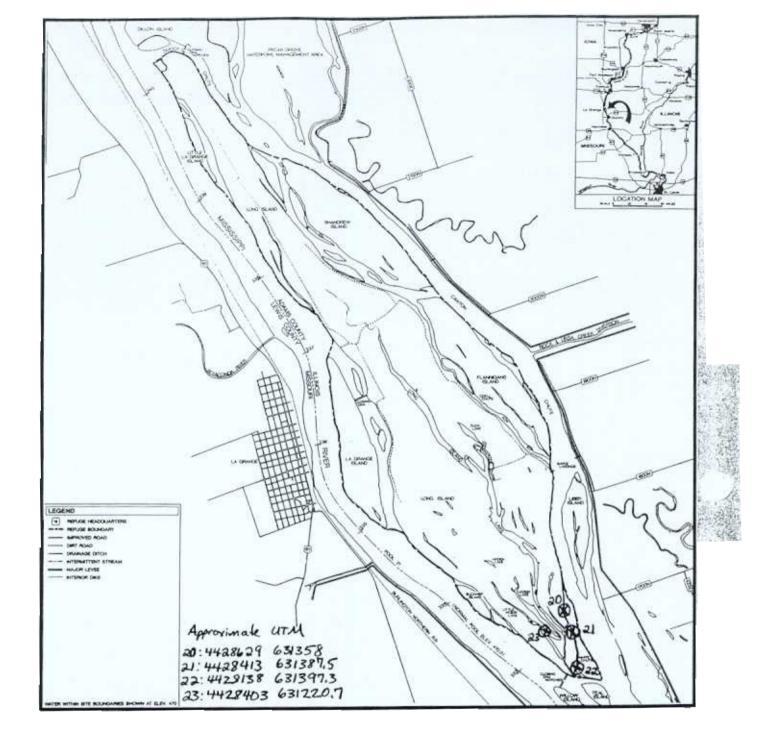




# KEITHSBURG DIVISION

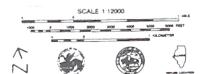
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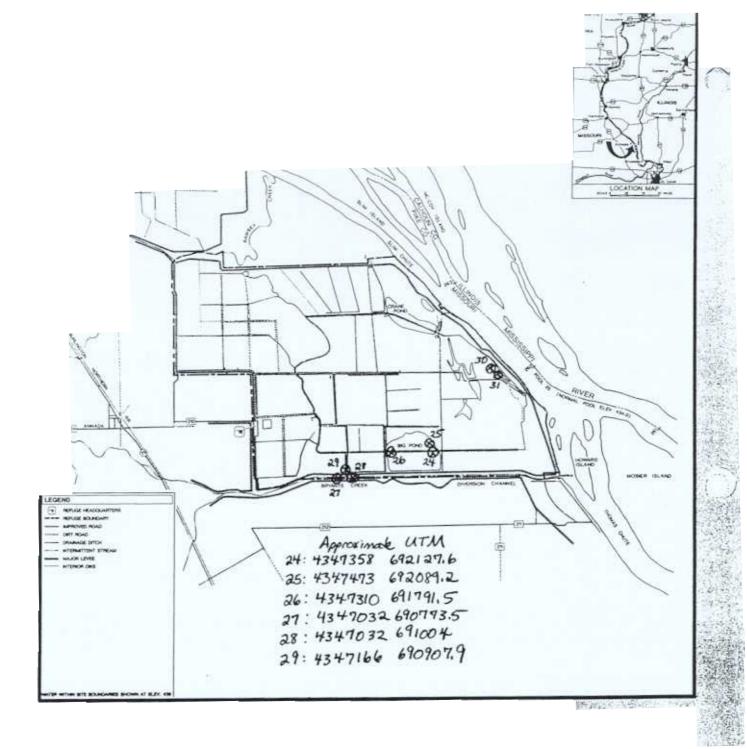




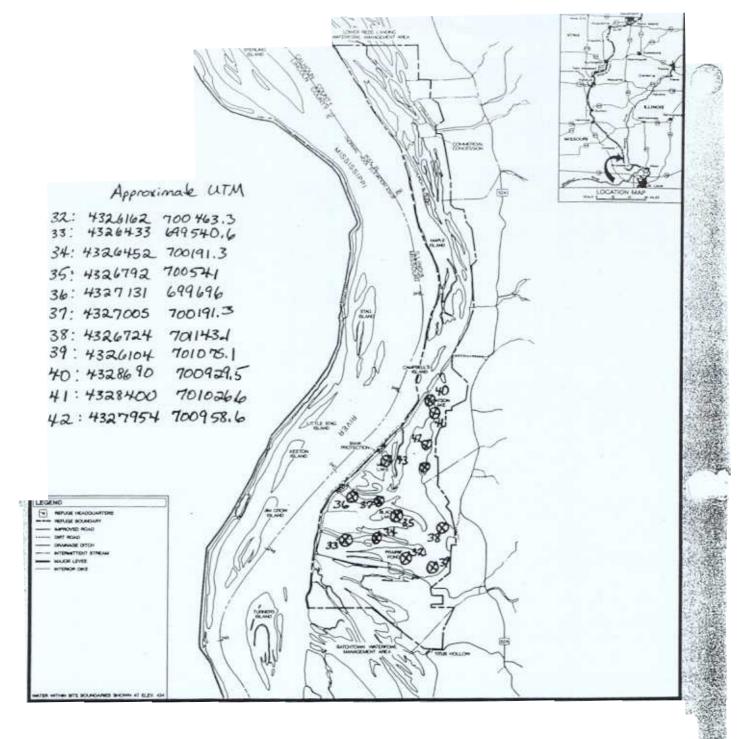
# **GARDNER DIVISION**

MARK TWAIN NATIONAL WILDLIFE REFUGE











# Appendix B Contaminant and Texture Data Records

Appendix B. Contaminant concentrations (micrograms per gram, dry weight) and texture data for the sampling sampling locations at Mark Twain National Widlife Refuge, 1992.

SITE	Reference No.	Zn	Pb	N	Cu	œ_	Cd	As	TOC	PO4	NH3-N	% CLAY	% SALT	% SAND
Skunk Slough	10	88.60	51.92	20.90	26.06	18.91	0.38	5.84	2.13	1100.00	55.00 70.00	50.62 47.59	44.54 42.95	4.84 9.46
	2a 3a	116.90 84.60	30.48 20.79	26.20 19.80	34.65 25.80	21. <b>84</b> 14.71	0.49 0.41	7.46 6.38	2.73 2.03		80.00	40.01	56.43	3.56
	16	95.20	25.12	21.10	30.30	16.84	0.43	6.55	2.65			50.80	44.83	4.37
	2b	92.30	32.00	21.70	31.46	16.30	0.45	7.30	2.78			49.57	45.51	4.92
	3b	78.80	20.74	18.00	25.20	14.13	0.40	6.00	2.00			36.61	60.18	3.31
(tig Timber Division	44	86.60	23.44	19.30	17.56	13.26	0.37	4.18 6.40	2.18 2.39	780.00	170.00 90.00	37.38 38.01	59.02 59.09	3.60 2.90
	6a 6a	107. <b>80</b> 33.20	25.43 46.48	25.40 17.40	21. <b>8</b> 0 6.31	17.24 6.75	0.48	3.06	0.17	700.00	50.00	7.39	4.80	88.01
	7a	106.20	35.32	26.10	22.40	22.56	0.45	5.87	1.79		80.00	24.51	42.68	32.51
	80	56.50	15.62	15.20	12.34	9.40	0.27	4.48	1.74	500.00		17.46	36.16	44.38
	9a	127.40	35.38	30.60	24.64	25.40	0.46	6.73	1.53	1000.00	62.00	35. <b>69</b>	56.15 55.67	6.16 11.71
	10a	112.70	23.46	24.80	23.36	18.45	0.46 0.45	6.96 6.73	2.17 1. <b>86</b>	860.00	59.00 140.00	32.62 36.66	53.85	9.49
	11a 12a	116.70 124.60	31.07 30.28	26.10 26.60	20.74 24.24	21.34 23.96	0.51	6.33	1.81		120.00	42.54	52.44	6.02
	4b	94.90	27.82	21.20	18.47	16.06	0.38	4.40	2.18			36.52	58.46	5.02
	5b	140.20	27.79	34.50	27.83	29.75	0.34	5.73	1.43			37.14	59.96	2.91
	60	31.20	0.00	13.90	5.92	7.83	0.00	3.04	0.49			6.83 22.91	3.80 42.82	90.37 34.27
	7b	96.10	20.55	24.90	20.81	50.52 11.02	0.44 0.29	0.50 4.29	1. <b>84</b> 2.06			20.37	39.39	40.24
	86 86	64.40 1,32.50	17.11 29.17	16.90 31.40	14.66 25.50	26.00	0.47	6.89	2.28			30.61	60.75	8.64
	106	124.80	33.15	28.70	23.10	22.38	0.47	8.15	2.30			41.06	45.46	12.86
	116	109.00	23.46	23.20	21.21	16.40	0.45	5.46	1.79			42.98	62.00	4.33
	12b	125.40	24.23	26.90	25.58	22.67	0.46	6.13	1.97		200 00	40.72 28.62	50.48 56.06	8.80 15.32
Keithsburg Division	13a	102.50	26.33	24.50	22.44	21.23	0.4 <b>5</b> 0. <b>65</b>	6.44 6.05	3.93 3.01		200.00 210.00	28.82 38.87	47.83	13.50
	14a 15a	122. <b>80</b> 76.20	26.43 17.86	26.00 20.60	26.93 20.54	21. <b>57</b> 24.27	0.68	6.64	3.78	2100.00	290.00	35.52	67.33	7.15
	16a	79.70	19.50	16.80	22.23	17.53	0.64	7.30	3.92	2.00.00	320.00	45.32	46.30	8.38
	17a	86.00	18.00	19.70	20.18	20.02	0.57	3.96	2.88	1900.00	190.00	31.85	81.96	6.20
	18a	74.60	23.31	15.70	22.06	12.90	0.58	6.22	3.83		210.00	39.78	49.78	10.44
	19e	67.40	20.46	16.10	20.70	13.36	0.50	5.63	3.93		200.00	32.65 26.42	50.96 64.21	16,40 9.55
	136 146	99.80 128.10	26.02 25.61	22.60 35.40	21.63 26.18	19.28 38.28	0.4 <b>5</b> 0.7 <b>6</b>	5.75 6.79	2.83 3.67			37.30	49.42	13.28
	156	73.80	19.29	16.10	21.61	15.74	0.61	6.00	4.31			37.30	55.28	7.42
	166	103.60	21.30	26.30	26.00	31.32	0.67	7.33	4.00			66. <del>6</del> 6	38.29	6.06
	176	99.90	20.50	21.90	23.60	22.30	0.50	6.35	2.38			28.03	63.06	8.91
	186	78.70	22.38	17.00	23.72	14.82	0.57	6.83	3.00 4.00			38.06 28.92	61.26 38.29	0.68 32.79
Gerdner Division	196 20a	60.40 18.00	16.32 9.26	1 <b>6.30</b> 0.00	16.74 10.34	16.66 3.60	0.43 0.30	6.57 2.93	1.58			12.17	57.28	30.65
Gerdner Division	21a	34.50	9.30	7.30	9.26	8.36	0.30	3.28	1.50			13.01	\$6.70	30.29
	220	50.70	11.29	10.70	11.54	10.06	0.37	3.82	1.55			16.81	71.60	11.59
	23e	86.70	18.70	16.70	16.10	14.25	0.50	8.10	1.32	1100.00		33.36	50.01	16.61
	20b	30.80	7.82	6.60	7.79	7.97	0. <b>26</b> 0. <b>32</b>	2.78 3.81	1.2 <b>3</b> 1.03			9.68 14. <b>89</b>	49.74 50.89	40.58 28.42
	21b 22b	55.00 56.40	10.10 . 13.15	12.10 11. <b>30</b>	11. <b>96</b> 12.77	11. <b>82</b> 10. <b>64</b>	0.41	4.24	1.07			19.19	72.74	8.07
	23b	56.30	14.34	14.30	11.80	17.80	0.80	5.76	1.62			22.58	65.44	21.98
Clarence Cannon Refuge	24a	63.50	23.53	25.90	21.43	23.26	0.49	3.34	2.82	800.00		67.13	36.67	7.20
•	26a	87.00	17.48	16.80	21.31	9.54	0.40	2.58	3.85		190.00	65.82 58.94	36.38 33.46	7.80 7.80
	260	55.30	17. <b>03</b> 13.07	13.00 17.40	21. <b>96</b> 17.22	7. <b>91</b> 19.01	0.36	3.06 4.86	2. <b>66</b> 0.67		79.00	45.83	50.16	4.01
	27a 28a	59.00 28.70	14.93	7.80	12.03	8.16	0.00	3.97	1.37		83.00	23.10	65.81	11.00
	290	44.40	12.46	15.70	17.51	8.86	0.00	3.65	1.31			23.30	80.06	7.63
	300	89.80	19.46	18.30	21.84	18.36	0.69	3.87	3.35		120.00	52.68	43,18	4.14
	31a	78.00	20.31	20.80	22.77	12.33	0.66	4.44	2.80	980.00	130.00	87.64 56.16	36.26 36.99	6.10 7.85
	24b 25b	97.80 83.80	27.80 19.83	24. <b>0</b> 0 1 <b>8.0</b> 0	26.85 23.83	20.08 12.73	0.51 0.44	3.34 2.67	3.71 3.11			\$5.50	36.98	6.62
	26b	55.70	18.18	9.50	22.77	6.20	0.37	3.04	2.17			59.15	35.01	5.84
	27b	84.60	15.76	22.00	16.58	17.56	0.00	4.89	1.30			35.19	80.39	4.42
	286	30.10	12.94	7.30	12.14	7.74	0.00	4.00	1.86			22.66	83.64	13.78
	306	77.80	17.93	20.70	21.91	11.26	0. <b>66</b> 0.77	3.06 3.06	3.30 3.48			50.83 55.62	43.62 38.21	8.66 6.17
Betchtown Division	31b 32a	75.40 83.00	17. <b>26</b> 25. <b>67</b>	19.70 19.90	21.4 <b>8</b> 21.42	10.77 23.93	0.77	8.00	2.39			52.28	38.00	9.63
BACKSONN CHARGO	33e	80.40	23.16	22.00	24.56	14.79	0.57	6.00	2.41	790.00	81.00	45.67	49.78	4.66
	344	103.90	20.74	29.00	24.84	28.16	0.52	5.64	2.26			44.66	60.02	5.33
	36a	145.60	25.26	37.40	31.17	35.67	0.61	7.27	1.86		110.00	52.33	42.29	5.38
	36a	106.20	23.23	29.50	28.08	22.80	0.55 0.51	7.18 6.09	2.47 2.4 <b>6</b>	1100.00 940.00	84.00	50.11 43.47	45.33 52.66	4. <b>56</b> 3.97
	37a 38a	99.30 115.00	20.99 26.46	44.30 33.50	25.92 27.01	46.00 27.4 <b>6</b>	0.50	7.20	1.90	3-10.00	<b>\$7.00</b>	48.16	49.08	2.76
	39a	60.10	18.36	18.60	17.16	9.73	0.48	4.11	2.22	730.00	58.00	31.57	60.77	7.66
	40a	47.50	11.67	14.70	10.42	10.73	0.23	2.70	1.26	620.00		13.82	53.86	32_33
	41a	33.00	10.50	13.10	8.40	10.04	0.20	3.41	0.98	900.00	40.00	14.97	81.96	3.07
	42a	82.10	20.62	23.60	22.12	20.51	0.4 <b>8</b> 0.43	6.84 6.34	1. <b>50</b> 2.19		130.00	33.36 38.68	63.24 55.76	3.41 6.67
	43a 32b	59.80 111.10	17.25 26.43	16.40 31.70	18.17 26.66	11.00 25.87	0.43	7.44	2.14		, 55.00	47.92	26.27	18.81
	336	100.80	22.54	28.10	25.46	24.76	0.56	5.51	2.40			46.32	48.33	6.32
	34b	126.30	23.06	30.90	29.60	26.37	0.52	5.41	2.36			46.86	50.64	2.80
	36b	132.30	25.54	36.40	30.98	33.51	0.62	7.36	2.67			54.07	42.22	3.71
	366	86.30	21.85	24.00	23.12	15.80	0.53	6.42	2.47			50.81 44.74	47.27 <b>53</b> .11	1. <b>82</b> 2.15
	37b	94.60	21.22	27.50	24.70 24.84	22.54 21.92	0. <b>58</b> 0.4 <b>9</b>	6. <b>68</b> 7.42	1. <b>66</b> 1. <b>69</b>			49.90	48.48	1.62
	38b <b>39</b> b	99.80 71.10	26.38 19.10	29.80 42.40	19.76	46.15	0.43	4.34	1.94			31.30	61.26	7.46
	406	38.40	10.71	12.00	9.26	7.31	0.21	2.36	0.84			14.24	49.59	36.17
	41b	35.00	11.06	12.50	6.63	9.97	0.21	4.14	0.86			16.54	81.87	1.59
	42b	61.70	19.66	18.00	19.46	12.61	0.44	6.00	1.61			32.90	63.23	3.87
	436	89.70	18.04	18.20	19.33	13.70	0.44	6.99	1.76			36.19	68.30	6.61

# Appendix C Macroinvertebrate Data Sheets

Reference number: Date processed:	0-1-1-5	9-25-92	0-1-1-6 9-29-92	9-29-92
Gastropods >1cm	R	R		
Gastropods <1cm	A	A	A	R
Sphaeriidae		3 anticulated	4 artimeted	6 articulared
Hexagenia sp.	5.	5	4	
Chironomidae <1cm				2
Chironomidae 1-4cm				3
Chironomidae >4cm			1	3
Heleidae				
	The second second second			
Reference number: Date processed:	4-1-3-5	4-1-2-7		
Date processed:				
Date processed:  Gastropods >1cm				
Date processed:  Gastropods >1cm  Gastropods <1cm		9-29-92		
Date processed:  Gastropods >1cm		9-29-92		
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae		9-29-92 12 1 anticulated		
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.	9.25-92	9-29-92 12 1 anticulated		
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.  Chironomidae <1cm	9.25-92	9-29-92 12 1 anticulated		
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm	9.25-92	9-29-92 12 1 anticulated		
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm  Chironomidae >4cm	9.25-92	9-29-92 12 1 anticulated		
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm  Chironomidae >4cm	9.25-92	9-29-92 12 1 anticulated		

Reference number: Date processed:	2-1-1+	2-1-1-	2-1-1-	
Gastropods >1cm				
Gastropods <1cm				56.40
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae				
Reference number:	2-1-2-7	2-1-2-6	5-1-5-8	
Date processed:	9-29-92	11-7-92	11-7-92	
Date processed: Gastropods >1cm	9-29-92	11-7-92	11-7-92	
Gastropods >1cm	4-29-92 A	11-7-92 R	11-7-92 A	
Gastropods >1cm Gastropods <1cm				
Gastropods >1cm Gastropods <1cm Sphaeriidae				
Gastropods >1cm Gastropods <1cm				
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp.	A			
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm	A	R.		
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm	A	R.		
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm Chironomidae >4cm	A I	3		
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm Chironomidae >4cm	A I	3		

Additional info, cite reference number:

2-1-2-6 lunger op of Nelsidae present

Reference number: Date processed:	2-1-4-7	2-1-4-6	2-1-4-8 11-7-92	
Gastropods >1cm				
Gastropods <1cm				
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm		1		
Chironomidae >4cm	3	3	1	
Heleidae			2	
heech		1		
shot, Pb				
Reference number: Date processed:			-	
Gastropods >1cm				
Gastropods <1cm				
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae	•			

Reference number:	2-2-1-	2-2-1-	2-2-1-	
Date processed:  Gastropods >1cm				
				70.0
Gastropods <1cm		The second secon		7782
Sphaeriidae				
Hexagenia sp.		1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1		
Chironomidae <1cm			18 V & 2 V &	
Chironomidae 1-4cm			u.	
Chironomidae >4cm				
Heleidae				
Reference number: Date processed:	2-2-2-7	2-2-2-6	2-2-2-	
Gastropods >1cm				
	_	A		
Gastropods <1cm		A -		
Gastropods <1cm Sphaeriidae		A		
Gastropods <1cm Sphaeriidae Hexagenia sp.		A		
Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm		A		
Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm	/	A 6		
Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm Chironomidae >4cm	3	A 6 1		
Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm	1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1 1	A 6 1		

Reference number: Date processed:	2-2-3-6	2-2-3-	2-2-3-8	
Gastropods >1cm	R			
Gastropods <1cm				
Sphaeriidae				
Hexagenia sp.				V.
Chironomidae <1cm				
Chironomidae 1-4cm			3	
Chironomidae >4cm	Q			
Heleidae	/		4	
Reference number:	2-2-4-	2-2-4-	2-2-4-	
Date processed:				
Date processed:  Gastropods >1cm				
Gastropods >1cm				
Gastropods >1cm Gastropods <1cm				
Gastropods >1cm Gastropods <1cm Sphaeriidae				
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp.				
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm				
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm				
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm Chironomidae >4cm				
Gastropods >1cm Gastropods <1cm Sphaeriidae Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm Chironomidae >4cm				

Reference number: Date processed:	3-1-1-6	3-1-1-4	3-1-1-	
Gastropods >1cm				
Gastropods <1cm	A	A		-11.
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae				
Worns	3	2		
Reference number: Date processed:	3-1-9-	3-1-2-	34-2-	
Gastropods >1cm				
Gastropods <1cm				
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae				
			Lagrana wing in the	

Additional info, cite reference number:

3-1-1-6 silk carls'- several

Reference number: Date processed:	3-2-1-4	3-2-1-	3-2-1	
Gastropods >1cm				
Gastropods <1cm	A			
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae		-3.		
Warms	4	3.1. -,;		
Reference number: Date processed:	3-2-2-5	3-2-3-6	3-2-2-	
Gastropods >1cm				
Gastropods <1cm	A	A		
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm		1		
Chironomidae >4cm				
Chironomidae >4cm Heleidae				

5-1-2-4	5-1-0-	5-1-9-	
			3
A			
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8			
6			
5-1-4-	5-1-4-	5-1-4-	
			and the state of t
	10-6-42 A 1 4 5 8	10-6-92 A 1 4 5 8	A  1  4  5  8  6

Reference number: Date processed:	5-1-6-	5-1-6-	5-1-6-	
Gastropods >1cm			4.	
Gastropods <1cm				
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae				
Reference number: Date processed:	5-1-8-5 9-25-92	5-1-8-4	5-1-8-	
Date processed:	9-25-92		5-1-8-	
Date processed:  Gastropods >1cm		5-1-8-4 11-9-92 R	5-1-8-	
Date processed:  Gastropods >1cm  Gastropods <1cm	9-25-92  R C Disortiulated	R	5-1-8-	
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae	9-25-92  R C Disorticulated	R	5-1-8-	
Date processed:  Gastropods >1cm  Gastropods <1cm	9-25-92  R C Disortiulated	R	5-1-8-	
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.	9-25-92  R C Disortiulated	R	5-1-8-	
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.  Chironomidae <1cm	9-25-92  R C Disortiulated	R	5-1-8-	
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm  Chironomidae >4cm	9-25-92  R C Disorticulated valves R	R	5-1-8-	
Date processed:  Gastropods >1cm  Gastropods <1cm  Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm	9-25-92  R C Disorticulated valves R	R C	5-1-8-	

Additional info, cite reference number:

"5-1-8-4 large 1-2 cm sp & Heleidal present

Rare= 3 or less; Common= 4 to 9; Abundant= 10 to 50; Dominant= greater than 50.

Reference number: Date processed:	5-1-10-	5-J-10 -	24-10-	
Gastropods >1cm		Walter Table		
Gastropods <1cm				
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae				
Reference number: Date processed:	5-1-12-	5-1-12-	5-1-12 -	4.3
Gastropods >1cm				
Gastropods <1cm				
Sphaeriidae				
Spliaerituae				1 2 1 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2 2
Hexagenia sp. Chironomidae <1cm				
Hexagenia sp.				
Hexagenia sp. Chironomidae <1cm				
Hexagenia sp. Chironomidae <1cm Chironomidae 1-4cm				
Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm  Chironomidae >4cm				
Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm  Chironomidae >4cm				

Reference number: Date processed:	6-0-1-	6-2-1	6-3-1	
Gastropods >1cm				
Gastropods <1cm				
Sphaeriidae				
Hexagenia sp.				
Chironomidae <1cm				
Chironomidae 1-4cm				
Chironomidae >4cm				
Heleidae				
Reference number: Date processed:	6-2-2	6-2-2	U-2-2-6 11-7-92	
Gastropods >1cm				
Gastropods <1cm				
Gastropods <1cm Sphaeriidae				
Sphaeriidae				
Sphaeriidae  Hexagenia sp.				
Sphaeriidae  Hexagenia sp.  Chironomidae <1cm				
Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm				
Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm  Chironomidae >4cm				
Sphaeriidae  Hexagenia sp.  Chironomidae <1cm  Chironomidae 1-4cm  Chironomidae >4cm				

# Appendix D Microtox® test Reports

FILE NAME: 3BWI 5-1- Sample Descript	1-57	1	TEST TIME:  Room temp. 22.3°C
:00pm, 2-10-93, Ke	ithsburg (W	inter), Detaile	
Procedure: SOLI Initial Concent Test Time: 5 m	ration : 98	684.2 ppm	Osmotic Adjustment: none Dilution Factor : 2 Concentration Units: ppm
NUMBER	It	CONC.	GAMMA
••••			0.0444
1	93.00	48.186	0.0466*
2	89.00	96.371	0.0936
3 4	73.00 61.00	192.743 385.485	0.3333 0.5956
5	50.00		0.3936
6	31.00	770.970 1541.941	2.1398#
7	14.00	3083.881	5.9524#
8	5.00	6167.763	18.4667#
9	3.00	12335.530	31.4444
10	3.00	24671.051	31.4444
11	1.00	49342.102	96.3333
12	1.00	98684.203	96.3333
CONTROL It'			.00
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·	EC	:50	•
: <u>=</u>	EC	<b>:50</b>	:
10 844444444		50 AAAAAAAAAAAAAAAAAA CONCENTRATION	

EC50 853.555 (95% CONFIDENCE RANGE 654.224 TO 1113.617)

Used for calculations \* Invalid games

FILE NAME: 2-1-3-3A.SPT				TEST DATE:	
Sam	ple Descript		Room tompo 30-8°C		
	12:55pm, 1	-29-93, Rou	nd Pond, Detail	ed Test, Dupe A	
Pro	cedure: SOLI	D-PHASE		Osmotic Adjustment: none	
Initial Concentration : 98684,2 ppm			Dilution Factor : 2		
Tes	t Time: 5 m	inutes		Concentration Units: ppm	
	Number	It	CONC.	GAMMA	
	•••••	98.00	40.104	0.001/+	
	1		48.186	0.0034*	
	2	104.00	96.371	-0.0545=	
	3	95.00	192.743	0.0351*	
	4	93.00	385.485	0.0573	
	5	98.00	770.970	0.0034*	
	6	91.00	1541.941	0.0806	
	7	91.00	3083.881	0.0806	
	8	105.00	6167.763	-0.0635*	
	9	66.00	12335.530	0.4899#	
	10	58.00	24671.051	0.6954#	
	11	35.00	49342.102	1.8095#	
	12	16.00	98684.203	5.1458#	
	CONTROL It'	s: 95.00	96.00 104	.00	
10	Desessassas	4444444444	44444444444444	44444444444444444444	
	*SLOPE - 1.	1558		•	
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	000	1024	CONCENTRATION		
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EC5027454.191 (95% CONFIDENCE RANGE:16731.852 TO 45047.773)

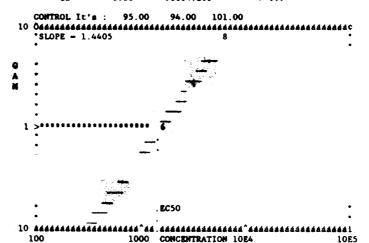
# Used for calculations \* Invalid gammas

TEST DATE: FILE NAME: 2-1-1-3B.SPT Sample Description:

3:15pm, 1-27-93, Round Pond, Detailed Test, Dupe B

Procedure: SOLID-PHASE Osmotic Adjustment: no Dilution Factor : 2 Concentration Units: 1 Initial Concentration: 98684.2 ppm Test Time: 5 minutes

NUMBER	It	CONC.	GANNIA
	• • • • • •		
1	97.00	48.186	-0.0034*
2	95.00	96.371	0.0175*
3	92.00	192.743	0.0507
4	78.00	385.485	0.2393
5	68.00	770.970	0.4216#
6	45.00	1541.941	1.1481#
7	25.00	3083.881	2.8667#
	10.00	6167.763	8.6667#
9	2.00	12335.530	47.3333
10	1.00	24671.051	95.6667
11	0.00	49342.102	> 999 +
12	0.00	98684.203	> 999 *



EC50 1417.029 (95% CONFIDENCE RANGE: 1284.887 TO 1562.760)

# Used for calculations \* Invalid gammas

```
FILE NAME: 2-1-1-3A.SPT
                                         TEST DATE:_
                                         TEST TIME:
Sample Description:
2:30; 1-27-93, Round Pond, Detailed Test, Dupe A. 21.7°C
Procedure: SOLID-PHASE
                                         Osmotic Adjustment: none
 Initial Concentration: 98684.2 ppm
                                         Dilution Factor : 2
Test Time: 5 minutes
                                         Concentration Units: ppm
       NUMBER
                  Ιt
                                CONC.
                                             GAMMA
                155.00
                              48.186
                                            -0.2968*
                             96.371
192.743
                116.00
                                            -0.0603*
         3
                107.00
                                             0.0187*
                100.00
87.00
72.00
35.00
                             385.485
                                            0.0900#
                             770.970
                                            0.2529#
                           1541.941
3083.881
6167.763
12335.530
                                            2.1143#
         8
                 15.00
                                            6.2667#
         q
                 3.00
                                           35.3333#
        10
                           24671.051
                 1.00
                                           108.0000*
        11
                 0.00
                           49342.102
                                           > 999 *
> 999 *
                 0.00
                           98684.203
 8
A
M
M
                               7
A
       .....................
                         6
                           EC50
1000 CONCENTRATION 10E4
```

EC50 1848.161 (95% CONFIDENCE RANGE: 1502.184 TO 2273.823)

# Used for calculations # Invalid gammas

```
FILE NAME: 0-1-2-3B.SPT
TEST DATE:
TEST TIME:
Sample Description:
1:10, 1-15-93, pool 19, Detailed Test, Dupe
Procedure: SOLID-PHASE
                                               Osmotic Adjustment: none
Dilution Factor: 2
Initial Concentration: 98684.2 ppm
Test Time: 5 minutes
                                                Concentration Units: ppm
       NUNBER
                    It
                                    CONC.
                                                    GANNIA
          1
                   63.00
                                  48.186
                                                    0.4656
                  117.00
                                   96.371
                                                   -0.2108*
                                192.743
385.485
770.970
1541.941
                   59.00
45.00
25.00
          3
                                                    0.5650#
                                                    1.0519#
          4
                                                    2.6933#
                   12.00
                                                    6.6944#
                               3083.881
6167.763
12335.530
24671.051
                    2.00
                                                   45.1667
                    4.00
2.00
1.00
          8
                                                   22.0833
                                                   45.1667
          10
                                                   91.3333
                    1.00
                               49342.102
                                                   91.3333
          12
                    0.00
                               98684.203
                                                   > 999 *
 "SLOPE - 1.2057
A
M
M
                                      .EC50
```

EC50 334.646 (95% CONFIDENCE RANGE 262.136 TO 427.212)

# Used for calculations \* Invalid gammas

FILE NAME: 0-1 Sample Descrip	tion:		TEST DATE: TEST TIME: Room Temp Off
2:15, 1-8	-93, pool	19, Detailed Test	:, Dupe B Temp. 24.6
Procedure: SOLI Initial Concent Test Time: 5 i	tration ·	98684.2 ppm	Osmotic Adjustment: none Dilution Factor : 2 Concentration Units: ppm
NUMBER	It	CONC.	GAMMA
1	106.00	48.186	0.000
2	92.00	96.371	-0.0031*
3	89.00	192.743	0.1486
4	77.00	385.485	0.1873#
. 5	59.00	770.970	0.3723#
6	37.00	1541.941	0.7910#
7.	25.00	3083.881	1.8559#
8	6.00	6167.763	3.2267#
9	3.00	12335.530	16.6111
10	2.00	24671.051	34.2222
11	2.00	49342.102	51.8333 51.8333
12 CONTROL It' 10 Öddddddddd	****	98684.203	> 999 *
CONTROL IE	s : 97.(	98684.203	> 999 *
CONTROL It' 10 Oddaddddddd "SLOPE - 1.0	s : 97.(	98684.203	> 999 *
CONTROL It's Oddaddaddaddaddaddaddaddaddaddaddaddadda	s : 97.(	98684.203	> 999 *
CONTROL It'.  10 04444444444  *SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 Oddaddddddd "SLOPE - 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It'. 10 04444444444 *SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	s : 97.(	98684.203	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	# : 97.( ####################################	98684.203 00 117.00 103. 1444444444444444444444444444444444444	> 999 *
CONTROL It' 10 0444444444 "SLOPE ~ 1.0	# : 97.( ####################################	98684.203	> 999 *
CONTROL IE: 10 0444444444 SLOPE - 1.6	# : 97.( ####################################	98684.203 DO 117.00 103.	> 999 *
CONTROL IE: 10 0444444444 SLOPE - 1.6	# : 97.( ####################################	98684.203 DO 117.00 103.	> 999 *

ECSO 950.852 (95% CONFIDENCE RANGE: 854.140 TO 1058,515)

FILE NAME: 0-1-2-3A.SPT TEST DATE: TEST TIME: Sample Description: Recommended to 12:10pm, 1-15-93, pool 19, Detailed Test, Dupe A Osmotic Adjustment: none Dilution Factor : 2 Procedure: SOLID-PHASE Initial Concentration: 98684.2 ppm Test Time: 5 minutes Concentration Units: ppm NUMBER CONC. GAHHA It 87.00 48.186 -0.0460\* 57.00 96.371 0.4561 70.00 192.743 0.1857# 60.00 48.00 27.00 385.485 770.970 1541.941 0.3833# 5 6 7 2.0741# 3083.881 6.5455# 40.5000 82.0000 2.00 6167.763 1.00 12335.530 9 10 0.00 24671.051 49342.102 > 999 \* > 999 11 98684.203 > 999 0.00 CONTROL It's : 94.00 79.00 76.00 \*SLOPE - 1.2714 G A N H . EC50 10 \* 1000 CONCENTRATION 10E4 100 10E5

EC50 813.897 (95% CONFIDENCE RANGE 649.133 TO 1020.483)

```
FILE NAME: 2-2-1-3A.SPT
                                          TEST DATE:
                                          TEST TIME:
   Sample Description:
                                           Roem Trong
                                                       21.4°C
3:00pm, 1-29-93, Little Denny Pond, Detailed Test, Dupe A
   Procedure: SOLID-PHASE
                                          Osmotic Adjustment: none
   Initial Concentration : 98684.2 ppm
                                          Dilution Factor: 2
   Test Time: 5 minutes
                                          Concentration Units: ppm
         NUMBER
                    Ιt
                                 CONC.
                                              GAMMA
                   91.00
                              48.186
96.371
192.743
                                             0.0659
                  100.00
                                             -0.0300*
                   91.00
88.00
                                             0.0659
                               385.485
                                              0.1023
                   84.00
71.00
47.00
                              770.970
                                              0.1548#
                              1541.941
                                             0.3662#
                              3083.881
                                             1.0638#
                   15.00
                              6167.763
                    9.00
                             12335.530
                                             9.7778#
                            24671.051
49342.102
           10
                    6.00
                                            15.1667
           11
                    4.00
                                             23.2500
           12
                             98684.203
                                             47.5000
      CONTROL It's :
                     95.00
                             97.00
                                     99.00
   G
  A
M
H
  A
                           6
                                . EC50
   1000 CONCENTRATION 10E4
     100
```

EC50 2667.740 (95% CONFIDENCE RANGE: 2096.438 TO 3394.727)

TEST DATE: TEST TIME: FILE NAME: 2-2-1-38.SPT Room Temp. Sample Description: :45pm, 1-29-93, Little Denny Pond, Detailed Test, Dupe B Procedure: SOLID-PHASE Osmotic Adjustment: none Initial Concentration: 98684.2 ppm Test Time: 5 minutes Dilution Factor : 2 Concentration Units: ppm NUMBER CONC. GAMMA 48.186 -0.2188\* 131.00 1 96.371 192.743 -0.1680\* 123.00 133.00 140.00 -0.2306\* 192.743 385.485 770.970 1541.941 3083.881 6167.763 12335.530 24671.051 -0.2690\* 118.00 105.00 80.00 51.00 21.00 12.00 -0.1328\* -0.0254\* 0.2792# 1.0065# 10 7.5278# 9.00 49342.102 10.3704# SLOPE - 1.3333 10 G A H H 10E6

EC50 6581.972 (95% CONFIDENCE RANGE: 3927.031 TO 11031.833)

FILE NAME: 2-2-	4-3A.SPT		TEST DATE:	
Sample Descript	ion:		Room temp. 21.40	
2:00pm, 1-29-93, Li	ttle Denny	Pond, Detailed		-
Procedure: SOLI Initial Concent	D-PHASE	3684.2 ppm	Osmotic Adjustment: Dilution Factor : 2	
Test Time: 5 m	inutes	•••	Concentration Units	
NUMBER	It	CONC.	GAMMA	
1	36.00	48.186	1.3611	
2	93.00	96.371	-0.0860*	
3	86.00	192.743	-0.0116*	
4	72.00	385.485	0.1806#	
5	49.00	770.970	0.7347	
6	27.00	1541.941	2.1481	
7	7.00	3083.881	11.1429	
8	1.00	6167.763	84.0000	
ğ.	1.00	12335.530	84.0000	
10	0.00	24671.051	> 999 *	
ii	0.00	49342.102	> 999 *	
12	1.00		84.0000	
CONTROL It'				
		CROTOX DATA REP		
FILE NAME: 2-2-	4-3A.SPT		TEST DATE:	
FILE NAME: 2-2- Sample Descript			TEST DATE:	
Sample Descript	ion:	Pond. Detailed 1	TEST TIME:	
Sample Descript :00pm, 1-29-93, L1 Procedure: SOLI	ion: ttle Denny		TEST TIME:	
Sample Descript :00pm, 1-29-93, L1 Procedure: SOLI	ion: ttle Denny		TEST TIME:  Test, Dupe A  Osmotic Adjustment:	
Sample Descript:00pm, 1-29-93, L1	ion: ttle Denny D-PHASE ration : 98		TEST TIME:	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLI Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration : 98 inutes	684.2 ppm CONC.	TEST TIME:  Fest, Dupe A  Osmotic Adjustment: Dilution Factor: 2	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration : 98 inutes	684.2 ppm	TEST TIME:  Test, Dupe A  Osmotic Adjustment: Dilution Factor : 2 Concentration Units:  GANNA	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLI Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration : 98 inutes It 36.00	684.2 ppm CONC.	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor : 2 Concentration Units:  GAMNA  1.3611	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLI Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration : 98 inutes It 	CONC	TEST TIME:  Test, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GANNA  1.3611 -0.0860*	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m: NUMBER 1 2 3	ion: ttle Denny D-PHASE ration: 98 inutes It 	CONC	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  CANNA  1.3611 -0.0860* -0.0116*	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m:  NUMBER 1 2 3 4	ion: ttle Denny D-PHASE ration : 98 inutes It 36.00 93.00 86.00 72.00	CONC	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  CANNA  1.3611 -0.0860* -0.0116* 0.1806*	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration : 98 inutes It 36.00 93.00 86.00 72.00 49.00	CONC. 48.186 96.371 192.743 385.485 770.970	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  CANNA  1.3611 -0.0860* -0.0116* 0.1806# 0.7347	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLI Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration: 98 inutes It 	CONC.  48.186 96.371 192.743 385.485 770.970 1541.941	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GANNA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration: 98 inutes It 36.00 93.00 86.00 72.00 49.00 27.00 7.00	CONC.  48.186 96.371 192.743 385.485 770.970 1541.941 3083.881	TEST TIME:  Test, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GAMMA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481 11.1429	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m:  NUMBER  1 2 3 4 5 6 7	ion: ttle Denny D-PHASE ration : 98 inutes It 36.00 93.00 86.00 72.00 49.00 27.00 7.00 7.00 1.00	CONC	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GAMMA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481 11.1429 84.0000	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m:  NUMBER  1 2 3 4 5 6 7 8 9	ion: ttle Denny D-PHASE ration : 98 inutes It 36.00 93.00 86.00 72.00 49.00 27.00 7.00 1.00 1.00	CONC	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  CANNA  1.3611 -0.0860* -0.0116* 0.1806# 0.7347 2.1481 11.1429 84.0000 84.0000	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLI Initial Concent: Test Time: 5 m NUMBER 1 2 3 4 5 6 7 8 9 10	10n: ttle Denny D-PHASE ration: 98 inutes It  36.00 93.00 86.00 72.00 49.00 27.00 7.00 1.00 0.00	CONC	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GANNA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481 11.1429 84.0000 84.0000 > 999 *	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m: NUMBER	ion: ttle Denny D-PHASE ration: 98 inutes It 36.00 93.00 86.00 72.00 49.00 27.00 7.00 1.00 1.00 0.00 0.00	CONC	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GANNA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481 11.1429 84.0000 84.0000 84.0000 > 999 * > 999 *	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m:  NUMBER	ion: ttle Denny D-PHASE ration: 98 inutes  It  36.00 93.00 86.00 72.00 49.00 27.00 7.00 1.00 1.00 0.00 0.00 0.00	CONC.  48.186 96.371 192.743 385.485 770.970 1541.941 3083.881 6167.763 12335.530 24671.051 49342.102 98684.203	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GANNA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481 11.1429 84.0000 84.0000 84.0000 > 9999 * > 999 * 84.0000	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m: NUMBER	Tettle Denny D-PHASE ration: 98 inutes  It  36.00 93.00 86.00 72.00 72.00 7.00 1.00 1.00 0.00 0.00 1.00 96.00	CONC	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GANNA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481 11.1429 84.0000 84.0000 > 999 * > 999 * 84.0000 00	
Sample Descript :00pm, 1-29-93, Li Procedure: SOLII Initial Concent: Test Time: 5 m:  NUMBER	Tettle Denny D-PHASE ration: 98 inutes  It  36.00 93.00 86.00 72.00 72.00 7.00 1.00 1.00 0.00 0.00 1.00 96.00	CONC.  48.186 96.371 192.743 385.485 770.970 1541.941 3083.881 6167.763 12335.530 24671.051 49342.102 98684.203	TEST TIME:  Cest, Dupe A  Osmotic Adjustment: Dilution Factor: 2 Concentration Units:  GANNA  1.3611 -0.0860* -0.0116* 0.1806* 0.7347 2.1481 11.1429 84.0000 84.0000 > 999 * > 999 * 84.0000 00	

TEST DATE: TEST TIME:

FILE NAME: 2-2-4-3A.SPT

Sample Description:

FILE NAME: 2-2-4-3B.SPT TEST DATE: TEST TIME: Sample Description: Room temp. 21.50C 2:45pm, 1-29-93, Little Denny Pond, Detailed Test, Dupe B Procedure: SOLID-PHASE Osmotic Adjustment: none Initial Concentration: 98684.2 ppm Test Time: 5 minutes Dilution Factor : 2 Concentration Units: ppm NUMBER Ιt CONC. GAMMA 1 92.00 48.186 0.0688 90.00 76.00 96.371 0.0926 192.743 0.2939 72.00 385.485 0.3657# 44.00 770.970 1.2348# 19.00 1541.941 4.1754# 3083.881 6167.763 12335.530 4.00 23.5833 0.00 > 999 0.00 > 999 10 0.00 24671.051 > 999 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999 CONTROL It's : 96.00 101.00 98.00 \*SLOPE - 1.7565 G A M H 1000 CONCENTRATION 10E4

EC50 683.532 (95% CONFIDENCE RANGE: 683.532 TO 683.532)

# Used for calculations \* Invalid gammas

10E5

MICROTOX DATA REPORT FILE NAME: 3-1-1-3A.SPT TEST DATE: TEST TIME: Sample Description: ROOM temp 31.4°C 4:00pm, 1-6-93, Keithsburg-lower, Detailed Test, Dupe A Procedure: SOLID-PHASE Osmotic Adjustment: none Initial Concentration: 98684.2 ppm Test Time: 5 minutes Dilution Factor: 2 Concentration Units: ppm NUMBER Ιt CONC. GAMMA 100.00 48.186 -0.0567\* 96.371 192,743 92.00 0.0254\* 94.00 0.0035\* 79.00 73.00 55.00 32.00 14.00 9.00 5.00 385.485 0.1941 5 770.970 0.2922# 1541.941 3083.881 6167.763 12335.530 0.7152# 1.9479# 5.7381# 9.4815 10 24671.051 17.8667 11 2.00 49342.102 46.1667 12 1.00 98684.203 93.3333 CONTROL It's : 97.00 82.00 104.00 \*SLOPE - 1.4332 8 M 7

EC50 1881.132 (95% CONFIDENCE RANGE: 1701.806 TO 2079.355)

Sample Descrip	otion:		TEST DATE:
			Room temp 31.50
20pm, 1-6-93, Ke		ver, Detailed Te	est, Dupe B
Procedure: SOI	ID-PHASE		Osmotic Adjustment: no
Initial Concer Test Time: 5	cration: 98	3684.2 ppm	Dilution Factor : 2
1000 1146.	mringces		Concentration Units: p
NUMBER	It	CONC.	GAMMA
1	103.00	48.186	0.0/52+
2	101.00	96.371	0.0453*
3	101.00	192.743	0.0660
4	92.00	385,485	0.0660
5	75.00	770.970	0.1703#
6	55.00	1541.941	0.4356# 0.9576#
7 ·	28.00	3083.881	2.8452#
8	10.00	6167.763	9.7667#
- 9	2.00	12335.530	52.8333
10	2.00	24671.051	52.8333
11	0.00	49342.102	> 999 *
12	0.00	98684.203	> 999 *
*SLOPE = 1	4391		######################################
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EC50 1403.855 (95% CONFIDENCE RANGE: 1206.177 TO 1633.929)

	LE NAME: 3-2-			TEST DATE:	
Sa	mple Descript	ion:		Roum Temp	25°€
1:00pm	, 1-8-93, Kei	thsburg-upp	er, Detailed Te	st, Dupe A	
In	ocedure: SOLI itial Concent	ration: 98	684.2 ppm	Osmotic Adjusts Dilution Factor	r : 2
Te	st Time: 5 m	inutes		Concentration l	Inits: ppm
	NUMBER	It	CONC.	GAMMA	
				*******	
	1	105.00	48.186	-0.0635*	
	2	94.00	96.371	0.0461*	
	3	95.00	192.743	0.0351*	
	4	80.00	385.485	0.2292#	
	5	66.00	770.970	0.4899#	
	6	49.00	1541.941	1.0068#	
	7	25.00	3083.881	2.9333	
	8	9.00	6167.763	9.9259	
	9	3.00	12335.530	31.7778	
	10	2.00	24671.051	48.1667	
	11	1.00	49342.102	97.3333	
	12	2.00	98684.203	48.1667	
	CONTROL It'			.00	
1			444444444444	<b>44444444</b> 4444444444444444444444444444	iaaaaaç
	*SLOPE - 1.	0677			•
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	•	4			•
	•	-	. EC50		•
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•	100	1000			10E5
		2000			LULJ

EC50 1522.536 (95% CONFIDENCE RANGE: 1289.541 TO 1797.630)

TEST DATE: TEST TIME: FILE NAME: 3-2-2-38.SPT Sample Description: Room temp 34.80C 1:30pm, 1-8-93, Keithsburg-upper, Detailed Test, Dupe B Procedure: SOLID-PHASE Osmotic Adjustment: none Initial Concentration: 98684.2 ppm Test Time: 5 minutes Dilution Factor : 2 Concentration Units: ppm CONC. GAIMIA NUMBER Ιt 48.186 96.371 192.743 385.485 100.00 0.0200\* 0.0099\* 101.00 95.00 85.00 0.0737 0.2000# 62.00 770.970 0.6452# 34.00 19.00 8.00 3.00 1541.941 2.0000# 4.3684 11.7500 3083.881 6167.763 12335.530 33,0000 10 2.00 24671.051 50.0000 49342.102 98684.203 109.00 101.00 101.0000\* 101.0000\* 1.00 1.00 96.00 12 CONTROL It's : \$ 10 044444444444444444444 \*SLOPE - 1.6610 G A M M A . EC50 1000 CONCENTRATION 1084 10E5

EC50 1011.776 (95% COMFIDENCE RANGE: 940.898 TO 1087.993)

FILE NAME: 4-1-1-3A.SPT TEST DATE: Sample Description:
1:30pm, 1-22-93, White Chute, Detailed Test, Dupé A Procedure: SOLID-PHASE Osmotic Adjustment: none Dilution Factor: 2 Initial Concentration: 98684.2 ppm Concentration Units: ppm Test Time: 5 minutes NUMBER CONC. GAMMA 1 86.00 48.186 0.0736 96.371 192.743 2 79.00 90.00 0.1688 0.0259\* 91.00 385.485 770.970 0.0147\* 39.00 1.3675# 84.00 1541.941 0.0992# 80.00 3083.881 0.1542# 8 47.00 6167.763 0.9645# 50.00 12335.530 10 30.00 24671.051 2.0778# 11 15.00 49342.102 5.1556# 12 7.00 98684.203 12.1905# CONTROL It's : 95.00 94.00 88.00 \*SLOPE - 0.7340 11 G A H H 10 . EC50 10E4 CONCENTRATION 10E5 10E6

EC50 8044.636 (95% CONFIDENCE RANGE: 2914.395 TO 22205.693)

```
FILE NAME: 4-1-1-3B.SPT
                                                TEST DATE:
                                                TEST TIME:
  Sample Description:
2:15pm, 1-22-93, White Chute, Detailed Test, Dupe B
  Procedure: SOLID-PHASE
                                               Osmotic Adjustment: none
Dilution Factor : 2
Concentration Units: ppm
  Initial Concentration: 98684.2 ppm
Test Time: 5 minutes
         NUMBER
                     Ιt
                                     CONC.
                                                   GAMMA
            1
                    94.00
                                   48.186
                                                   0.0638
                                 96.371
192.743
                    95.00
                                                   0.0526
            3
                    98.00
90.00
90.00
                                                   0.0204*
                                  385.485
                                                   0.1111
                                  770.970
                                                   0.1111
                    93.00
                                1541.941
                                                   0.0753
                    82.00
                               3083.881
6167.763
12335.530
24671.051
                                                   0.2195
                    79.00
                                                   0.2658#
0.8519#
                   54.00
33.00
18.00
11.00
           10
                                                   2.0303#
                               49342.102
                                                   4.5556#
          12
                               98684.203
     CONTROL It's :
                                                   8.0909
                       95.00
                               108.00
                                        97.00
  *SLOPE - 1.3550
 G
 A
M
M
 Α
                              .EC50
10E4 CONCENTRATION 10E5
```

EC5015236.967 (95% CONFIDENCE RANGE:12339.940 TO 18814.123)

TEST DATE: FILE NAME: 4-1-3-3A.SPT TEST TIME: Room temp. 22./°C 12:10, 1-22-93, White Chute, Detailed Test, Dupe A Sample Description: Osmotic Adjustment: none Procedure: SOLID-PHASE Initial Concentration: 98684.2 ppm Test Time: 5 minutes Dilution Factor : 2 Concentration Units: ppm GAMMA NUMBER Ιt CONC. 96.00 86.00 48.186 -0.0208\* 96.371 0.0930 91.00 192.743 0.0330= 385.485 770.970 1541.941 3083.881 0.1463 0.3429 82.00 70.00 62.00 40.00 0.5161# 1.3500# 20.00 6167.763 3.7000# 12.4286 46.0000 93.0000 7.00 12335.530 24671.051 49342.102 98684.203 10 2.00 1.00 11 93.0000 1.00 97.00 12 10 \*SLOPE - 1.4208 G H EC50 100 1000 CONCENTRATION LOEA 10E5

EC50 2469.598 (95% CONFIDENCE RANGE: 2208.433 TO 2761.648)

Sample Descrip	tion:		TEST TIME:	
1:00, 1-2	2-93, White	Chute, Detailed	Roontemp. 3 Test, Dupe B	2.2°C
Procedure: SOL	ID-PHASE		Osmotic Adjust	ment: none
Initial Concent Test Time: 5 :	tration: 98	684.2 ppm	Dilution Facto	r : 2
rest lime; ) i	nutes		Concentration	Units: ppm
NUMBER	It	CONC.	GAMMA	
1	130.00	48.186		
2	114.00	96.371	-0.1128*	
3	116.00	192,743	0.0117*	
4	114.00	385.485	-0.0057*	
5	72.00		0.0117*	
6	0.00	770.970	0.6019#	
7	70.00	1541.941 3083.881	> 999 *	
8	49.00		0.6476#	
9	22.00	6167.763 12335.530	1.3537#	
10	7.00	24671.051	4.2424#	
		240/1.031	15.4762#	
11	4.00	49342 102	27 02224	
11 12 CONTROL It' 10 044444444444444444444444444444444444	444444444	49342.102 98684.203 126.00 124. 4 6 44444444444	27.8333# 37.4444 00 444444444 10 44	11 444¢
12 CONTROL It' 10 044444444 *SLOPE = 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	11 ssac
12 CONTROL It' 10 044444444 'SLOPE - 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	11 444¢
12 CONTROL IE' 10 0444444444 *SLOPE = 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	11 444¢
12 CONTROL It' 10 0444444444 *SLOPE = 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	11 444¢
12 CONTROL It' 10 0444444444 *SLOPE = 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	11 <b>444</b> ¢
12 CONTROL IE' 10 044444444444444444444444444444444444	3.00 s: 96.00	98684,203 126,00 124	37.4444	11 444¢
12 CONTROL It' 10 0444444444 *SLOPE = 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	11 4440
12 CONTROL IE' 10 044444444444444444444444444444444444	3.00 s: 96.00	98684,203 126,00 124	37.4444	
12 CONTROL It' 10 044444444 *SLOPE - 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	
12 CONTROL It' 10 044444444 *SLOPE - 0.	3.00 s: 96.00	98684,203 126,00 124	37.4444	
12 CONTROL IE' 10 044444444444444444444444444444444444	3.00 s: 96.00	98684,203 126,00 124	37.4444	
12 CONTROL IE' 10 044444444444444444444444444444444444	3.00 s: 96.00	98684,203 126,00 124	37.4444	
12 CONTROL IE' 10 044444444444444444444444444444444444	3.00 s: 96.00	98684,203 126,00 124	37.4444	
12 CONTROL It' 10 044444444444444444444444444444444444	3.00 s: 96.00 44444444444 2535	98684.203 126.00 124. 4 6 444444444444444444444444444444444	37.4444 00 144444444 10 44	
12 CONTROL It' 10 044444444444444444444444444444444444	3.00 s: 96.00 44444444444 2535	98684.203 126.00 124. 4 6 444444444444444444444444444444444	37.4444	• • • • • • • • • • • • • • • • • • •

EC50 544.984 (95% CONFIDENCE RANGE: 12.458 TO 23840.254)

<sup>#</sup> Used for calculations
\* Invalid gammas

Sample	AME: 5-1-: Descript:	1-3A.SPT		TEST DATE: TEST TIME:  Room temp. 23.5	_ _ 
Ор <b>м</b> , 2-	3-93, Bate	chtown I ar	d II, Detailed	Test, Dupe A	<i>ا</i> در
Initia	ure: SOLI 1 Concent 1me: 5 m	ration : 96	684.2 ppm	Osmotic Adjustment: Dilution Factor : 2 Concentration Units	
	NUMBER	It	CONC.	GAMMA	
	1	90.00	48.186	0.0026	
	2	79.00	96.371	0.0926	
	3	73.00	192.743	0.2447	
	4	50.00		0.3470#	
	5	22.00	385.485 770.970	0.9667#	
	6	6.00		3.4697#	
	7	1.00	1541.941	15.3889#	
	8	1.00	3083.881	97.3333	
	9	0.00	6167.763	97.3333	
	10	0.00	12335.530	> 999 *	
	11	0.00	24671.051	> 999 *	
	12		49342.102	> 999 *	
10 Öá	NTROL It's	444444444		> 999 + .00 44444 6 ###############################	ic
10 Öá	NTROL It's	s : 96.00	98.00 101		ic •
10 Ö4.	NTROL It's	s : 96.00	98.00 101	.00	ic .
10 Ö4.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4.	NTROL It's	s : 96.00	98.00 101	.00	ic .
10 Ö4.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Öá	NTROL It's	s : 96.00	98.00 101	.00	•
10 Öá	NTROL It's	s : 96.00	98.00 101	.00	ic
10 Ö4SG. AHHA.	NTROL It's	s : 96.00	98.00 101	.00	ic
10 Ö4SG. AHH.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. AHHA.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. AHH.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. AHHA.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. AHHA.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. AHHA.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. A. H. H. A.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. A. H. H. A.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Ö4SG. A. H. H. A.	NTROL It's	s : 96.00	98.00 101	.00	•
10 Öá. 'S' 'S' 'G' A 'H' H 'A '  1 >* ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' '	NTROL It'	s: 96.00 444444444 8256	98.00 101	.00	• • • • • • • • • • • • • • • • • • •
10 Öá. 'S' 'S' 'G' A 'H' H 'A '  1 >* ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' '	NTROL It'	s : 96.00	98.00 101	.00	• • • • • • • • • • • • • • • • • • •

EC50 368.185 (95% CONFIDENCE RANGE: 296.380 TO 457.388)

<sup>#</sup> Used for calculations \* Invalid gammas

FILE NAME: 5-1-	1 • 3B . SPT		TEST DATE:
			TEST TIME:
Sample Descript	ion:		Room Temp: 23.4°C.
:00pm, 2-3-93, Bat	chtown I ar	d II, Detailed	Test, Dupe B
Procedure: SOLI	D-PHASE		Osmotic Adjustment: none
Initial Concent Test Time: 5 m		1684.2 ppm	Dilution Factor : 2 Concentration Units: ppe
NUMBER	It	CONC.	GAMMA
1	96.00	48.186	0.1181
2	83,00	96.371	0.2932
3	76.00	192,743	0.4123
4	57.00	385.485	0.8830w
5	31.00	770.970	2.4624#
6	12.00	1541.941	7.9444#
7	4.00	3083.881	25.8333#
8	2.00	6167.763	52.6667
9	1.00	12335.530	106.3333*
10	1.00	24671.051	106.3333*
11	1.00	49342.102	106.3333*
12	0.00	98684.203	> 999 *
**CONTROL It's  10 044444444444  **SLOPE - 1.4	******		0.00 
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	. EC50		•
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10 44444444444 100	14.44444^4 1000		444 <sup>4</sup> 44444444444444444444441 10E4 10E5

EC50 428.209 (95% CONFIDENCE RANGE. 379.531 TO 483.129)

FILE NAME: 5-1-4-3A.SPT

TEST DATE:

Sample Description:

Room temp. 23.5 C

2:25pm, 2-3-93, Batchtown I and II, Detailed Test, Dupe A

Procedure: SOLID-PHASE Initial Concentration: 98684.2 ppm Test Time: 5 minutes

684.2 ppm Dilui

Osmotic Adjustment: none Dilution Factor : 2 Concentration Units: ppm

NUMBER	It	CONC.	GAMMA
	*******	•••••	
1	77.00	48.186	0.2251
2	76.00	96.371	0.2412#
3	68.00	192.743	0.3873#
4	39.00	385.485	1.4188#
5	25.00	770.970	2.7733#
6	16.00	1541.941	4.8958#
7	5.00	3083.881	17.8667#
	1.00	6167.763	93.3333
9	1.00	12335.530	93.3333
10	0.00	24671.051	> 999 *
11	0.00	49342.102	> 999 *
12	0.00	98684.203	> 999 *

	*SLOPE - 1.2286				·
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ECSO 343.202 (95% CONFIDENCE RANGE: 272.486 TO 432.271)

FILE NAME: 5-1-4	TEST DATE: TEST TIME:			
Sample Descript:			Room temp. 234	
P <sup>P</sup> 2-3-93, Batchtown I and II, Detailed Te			st, Dupe B	
Procedure: SOLII Initial Concent: Test Time: 5 m	ration : 98	6684.2 ppm	Osmotic Adjustment: Dilution Factor : 1 Concentration Units	
NUMBER	It	CONC.	GAMMA	
1	95.00	48.186	-0.0175*	
2	85.00	96.371	0.0980#	
. 3	67.00	192.743	0.3930#	
4	57.00	385.485	0.6374#	
5	26.00	770.970	2.5897#	
6	19.00	1541.941	3.9123#	
7	7.00	3083.881	12.3333#	
8	3.00	6167.763	30.1111	
9	2.00	12335.530	45.6667	
10	1.00	24671.051	92.3333	
11	1.00	49342.102	92.3333	
12	0.00	98684.203	> 999 *	
SLOPE - 1.3	384		44444444444444444444444	
1 >**************				
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•	. EC50		444^4444444444444	

EC50 475.297 (95% CONFIDENCE RANGE 380.706 TO 593.389)

5-1-7-3A -PHASE ation: 9.8 nutes	68 <b>t</b>	Smotic Adjustment: no Dilution Factor: 2
stion : 9.8 nutes	68 %	Dilution Factor : 2
It		Concentration Units: 4
•••••	CONC.	GAMMA
131.00 126.00 116.00 105.00 88.00 52.00 22.00 7.00 1.00 1.00	0.0048 0.0096 0.0193 0.0385 0.0771 0.1542 0.3084 0.6168 1.2335 2.4670 4.9340 9.8680 136.00 136.444444444444444444444444444444444444	-0.0611* -0.0238* 0.0603 0.1714 0.3977# 1.3654# 4.5909# 16.5714# 122.0000* 122.0000* 122.0000* 122.0000*
	<u>-</u>	a a 5 0 0
	126.00 116.00 105.00 88.00 52.00 22.00 7.00 1.00 1.00 1.00 297.00 1.00 297.00	126.00 0.0096 116.00 0.0193 105.00 0.0185 88.00 0.0771 52.00 0.1542 22.00 0.3084 7.00 0.6168 1.00 1.2335 1.00 2.4670 1.00 4.9340 1.00 9.8680 97.00 136.00 136.

0.1297 (95% CONFIDENCE RANGE: 0.1254 TO 0.1340

FILE NAME: 5-1-	7 - 3A . SPT		TEST DATE:	
Sample Descript	ion:		Roon temp.	22.44
2:00pm, 1-13-93, Ba	tchtown I a	nd II, Detailed		
Procedure: SOLI Initial Concent Test Time: 5 m	ration : 98	684.2 ppm	Osmotic Adjust Dilution Facto Concentration	r : 2
NUMBER	lt	CONC.	GANNA	
1 2 3 4 5 6 7 8 9 10 11 12 CONTROL IC' 10 Odddddddddddddddddddddddddddddddddddd	4444444444		-0.0611* -0.0238* 0.0603 0.1714 0.3977# 1.3654# 4.5909# 16.5714# 122.0000* 122.0000* 122.0000* 122.0000*	444444C • • •
; 1 >************************************		. EC50		:
100	1000	.44444444444444 CONCENTRATION		444441 10E5

EC50 1296.576 (95% CONFIDENCE RANGE: 1255.075 TO 1339.449)

	m.	CRUTUX DATA REP	ORT	
FILE NAME: 5-1-	7-3B.SPT		TEST DATE:	
Sample Descript		<del>,</del>		
3:00mm 1 12 02 n-			_ Room temp 22.5	
3:00pm, 1-13-93, Ba	tentown 1 a	nd II, Detailed	Test, Dupe B	
Procedure: SOLI			Osmotic Adjustment	t: noi
Initial Concent	ration : 98	684.2 ррш	Dilution Factor :	2
Test Time: 5 m	inutes		Concentration Uni	s: p
NUMBER	It	CONC.	GAMMA	
1	89.00	48.186	0.0524	
2	80.00	96.371	0.0324	
3	80.00	192.743	0.1708#	
4	67.00	385.485	0.3980#	
5	53.00	770.970	0.7673#	
6	32.00	1541.941	1.9271#	
7	12.00	3083.881	6.8056#	
8	5.00	6167.763	17.7333#	
9	2.00	12335.530	45.8333	
10	1.00	24671.051	92.6667	
11	1.00	49342.102	92.6667	
12	1.00	98684.203	92.6667	
CONTROL It'	s: 96.00	89.00 96	. 00	
10 044444444	*444444444	8 ******	<b>444</b> 4444444444444444	ááC
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10 444444444	44444 114		*********	•
100	88 8.85888=== ^^^^	CONCENTRATION		
200	1000	CONCENTRATION	IUE4	10E5

EC50 802.832 (95% CONFIDENCE RANGE 678.868 TO 949.432)

# Used for calculation

11.	CONOTON DATA KEN	OKI
		TEST DATE:
ion:		
h town I ar	d II. Detailed (	Room Temp: 21,9°C
	, Document	resc, supe A
ration : 98	684.2 ppm	Osmotic Adjustment: none Dilution Factor : 2 Concentration Units: ppm
It	CONC.	GAMMA
98.00	48 186	-0.0544*
90.00		0.0296*
95.00		-0.0246*
83.00		0.1165
78.00	770.970	0.1880#
	1541.941	0.5444#
	3083.881	1.3761#
		4.7917#
	12335.530	29.8889
	446/1.051	45.3333
		91.6667
: 95.00	90 00 03	91.6667
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1000	CONCENTRATION 1	.0E4 10E5
	11-A.SPT ion: h town I ar D-PHASE ration: 98 inutes  It 98.00 99.00 95.00 83.00 78.00 60.00 39.00 16.00 3.00 2.00 1.00 1.00 1.00 2.00 3.00 2.00 3.00 2.00 3.00 2.00 3.00 2.00 3.00 2.00 3.00 2.00 3.00 2.00 3.00 2.00 3.00 3	11-A.SPT  ion: h town I and II, Detailed of the country of the cou

EC50 2324.261 (95% CONFIDENCE RANGE: 2027.790 TO 2664.076)

40pm, 1-15-93, Batch town I and II, Detailed Test, Dupe B  Procedure: SOLID-PHASE Osmotic Adjustment: Initial Concentration: 98684.2 ppm Dilution Factor: 2		mionotom baim al	
### ### ##############################	FILE NAME: 5-1-11-B	SPT	
Procedure: SOLID-PHASE	Sample Description:		Room temp. 21.900
Initial Concentration: 98684.2 ppm	pm, 1-15-93, Batch	town I and II, Detail	• • • •
Initial Concentration: 98684.2 ppm Test Time: 5 minutes  NUMBER It CONC. GAMMA  1 105.00 48.186 -0.2825* 2 103.00 96.371 -0.2686* 3 100.00 192.743 -0.2467* 4 106.00 385.485 -0.2893* 5 86.00 770.970 -0.1240* 6 73.00 1541.941 0.0320* 7 45.00 3083.881 0.6741* 8 22.00 6167.763 2.4242* 9 6.00 12335.530 11.5556* 10 2.00 24671.051 36.6667* 11 1.00 49342.102 74.3333 12 1.00 98684.203 74.3333 CONTROL It's: 97.00 49.00 80.00  10 044444444444444444444444444444444	Procedure: SOLID-PH	ASE	Osmotic Adjustment: non
1 105.00 48.186 -0.2825* 2 103.00 96.371 -0.2686* 3 100.00 192.743 -0.2467* 4 106.00 385.485 -0.2893* 5 86.00 770.970 -0.1240* 6 73.00 1541.941 0.0320* 7 45.00 3083.881 0.6741* 8 22.00 6167.763 2.4242* 9 6.00 12335.530 11.5556* 10 2.00 24671.051 36.6667* 11 1.00 49342.102 74.3333 12 1.00 98684.203 74.3333 CONTROL It's: 97.00 49.00 80.00 10 0444444444444444444444444444444444			
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8 22.00 6167.763 2.4242# 9 6.00 12335.530 11.5556# 10 2.00 24671.051 36.6667# 11 1.00 49342.102 74.3333 12 1.00 98684.203 74.3333  CONTROL It's: 97.00 49.00 80.00 10 0444444444444444444444444444444444			0.0320*
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EC50 3788.811 (95% CONFIDENCE RANGE: 3134.043 TO 4580.374)

Sample Descript	3-3A.SPT	Pond Details	TEST DATE: TEST TIME: Room Temp.: 01.90
Procedure: SOLI Initial Concent Test Time: 5 m	D-PHASE	Pond, Detailed	Osmotic Adjustment: none Dilution Factor : 2 Concentration Units: ppm
NUMBER	It	CONC.	GAMMA
1	73.00	48.186	0.1644
2	73.00	96.371	0.1644
3	72.00	192.743	0.1806
4	65.00	385.485	0.1808
5	56.00	770.970	
6	37.00	1541.941	0.5179#
ž	21.00		1.2973#
8	5.00	3083.881	3.0476#
9		6167.763	16.0000
10	3.00	12335.530	27.3333
	3.00	24671.051	27.3333
11 12	0.00 0.00	49342.102 98684.203	> 999 *
			> 999 *
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EC50 1279.279 (95% CONFIDENCE RANGE: 1092.498 TO 1497.993)

FILE NAME: 6-1-3-3B.SPT	TEST DATE:
Sample Description: 2:00pm, 2-12-93, Big Pond,	TEST TIME:  Room temp. 21.9°C  Detailed Test, Dupe B

Procedure: SOLID-PHASE
Initial Concentration: 98684.2 ppm Dilution Factor: 2
Test Time: 5 minutes Concentration Units: ppm

NUMBER	It	CONC.	GAMMA
• • • • • •	• • • • • • •		
1	0.00	48.186	> 999 *
2	139.00	96.371	-0.7650*
3	0.00	192.743	> 999 *
4	0.00	385.485	> 999 *
5	0.00	770.970	> 999 *
6	115.00	1541.941	-0.7159*
7	83.00	3083.881	-0.6064*
8	15.00	6167.763	1.1778
9	5.00	12335.530	5.5333
10	2.00	24671.051	15.3333
11	1.00	49342.102	31.6667
12	0.00	98684.203	> 999 *

CONTROL It's: 98.00 0.00 0.00
......NO TOXICITY AT CONCENTRATIONS TESTED
\* Invalid gammas

TEST DATE:

FILE NAME: 6-1-4-3A.SPT

TEST TIME: le Description:
Recm tomp. 32.0°C
2:15pm, 2-10-93, Big Pond, Detailed Test, Dupe A Sample Description: Procedure: SOLID-PHASE Osmotic Adjustment: none Initial Concentration: 98684.2 ppm Test Time: 5 minutes Dilution Factor : 2 Concentration Units: ppm NUMBER CONC. Ιt GAMMA 1 111.00 48.186 -0.0991\* 100.00 96.371 0.0000\* 3 94.00 192.743 0.0638 385.485 770.970 1541.941 3083.881 6167.763 90.00 0.1111 80.00 0.2500 64.00 0.5625# 32.00 2.1250# 19.0000# 8 5.00 9 1.00 12335.530 99.0000# 10 0.00 24671.051 > 999 \* 11 0.00 49342.102 > 999 \* 12 0.00 98684.203 > 999 101.00 104.00 CONTROL It's : 95.00 \*SLOPE - 2.5539 G A M M 1 >4 . EC50 100 1000 CONCENTRATION 10E4 10E5

EC50 2060.982 (95% CONFIDENCE RANGE: 1539.959 TO 2758.287)

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units: ¡
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EC50 2129.088 (95% CONFIDENCE RANGE: 1821.125 TO 2489.130)

Sample Description: Room 1  15pm, 2-12-93, Bryrants Creek, Detailed Test, Dupe A  Procedure: SOLID-PHASE Osmotic Initial Concentration: 98684.2 ppm Dilution	E:		
15pm, 2-12-93, Bryrants Creek, Deteiled Test, Dupe A  Procedure: SOLID-PHASE Initial Concentration: 98684.2 ppm Dilution Test Time: 5 minutes Conc. GAMM  1 82.00 48.186 0.15 2 100.00 96.371 -0.05 3 117.00 192.743 -0.19 4 97.00 385.485 -0.02 5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00  10 044444444444444444444444444444444	Room Temp! 20.7°		
Initial Concentration: 98684.2 ppm Dilution Test Time: 5 minutes Concentration: 1 82.00 48.186 0.15 2 100.00 96.371 -0.05 3 117.00 192.743 -0.19 4 97.00 385.485 -0.02 5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444	J .		
NUMBER It CONC. GAMM  1 82.00 48.186 0.15 2 100.00 96.371 -0.05 3 117.00 192.743 -0.19 4 97.00 385.485 -0.02 5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444	Adjustment: no		
NUMBER It CONC. GAMM  1 82.00 48.186 0.15 2 100.00 96.371 -0.05 3 117.00 192.743 -0.19 4 97.00 385.485 -0.02 5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.266 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0434444444444444444444444444444444444	Factor : 2		
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1 82.00 48.186 0.15 2 100.00 96.371 -0.05 3 117.00 192.743 -0.19 4 97.00 385.485 -0.02 5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444			
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3 117.00 192.743 -0.19 4 97.00 385.485 -0.02 5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444			
4 97.00 385.485 -0.02 5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444			
5 90.00 770.970 0.05 6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444			
6 73.00 1541.941 0.29 7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444	-		
7 49.00 3083.881 0.93 8 29.00 6167.763 2.26 9 4.00 12335.530 22.66 10 1.00 24671.051 93.66 11 0.00 49342.102 > 999 12 0.00 98684.203 > 999  CONTROL It's: 95.00 93.00 96.00 10 0444444444444444444444444444444444			
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EC50 3140.505 (95% CONFIDENCE RANGE: 2563.958 TO 3846.698)

		***	ORUTOR DATE REE	<b>~</b>	
FILE	E NAME: 6-2-	1-3B.SPT		TEST DATE:	_
				TEST TIME:	
Samp	ole Descript	ion:		Rain Temp. : 5	21.9 0
1:15pm,	2-12-93, Br	yrants Cree	k, Detailed Tes		
Proc	edure: SOLI	D-PHASE		Osmotic Adjustme	nt: none
	ial Concent		684.2 ppm	Dilution Factor	: 2
Test	:Time: 5 m	inutes		Concentration Un	its: ppm
	NUMBER	It	COMC.	GANNA	
	1	113.00	48.186	-0.2153*	
	2	108.00	96.371	-0.2133* -0.1790*	
	3	104.00	192.743	-0.1474*	
	4	111.00	_		
	5		385.485	-0.2012 <del>*</del>	
	6	0.00	770.970	> 999 ±	
	7	80.00	1541.941	0.1083#	
	-	49.00	3083.881	0.8095#	
	8	30.00	6167.763	1.9556#	
	9	7.00	12335.530	11.6667#	
	10	2.00	24671.051	43.3333#	
	11	1.00	49342.102	87.6667#	
	12	1.00	98684.203	87.6667	
10	CONTROL It' O444444444 *SLOPE - 0.	444444 5 44		'.00 4466 9 466 10 66 11	4440
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EC50 842.851 (95% CONFIDENCE RANGE 19.604 TO 36237.098)

Sample Descript :15pm, 2-12-93, Br Procedure: SOLI Initial Concent			Room Temp .:	
Procedure: SOLI	•	ek. Detailed Te	st Dune A	જા. ၂ . (
Initial Concent	D-PHASE			
	ration . Qf	1686 2	Osmotic Adjust	ment: none
Test Time: 5 m	inutes	OUT. E ppus	Dilution Facto	r : 2
			Concentration	Units: ppa
NUMBER	It	CONC.	GAMMA	
		• • • • • • • • • • • • • • • • • • • •	•••••	
1	116.00	48.186	-0.0776*	
2	110.00	96.371	-0.0273*	
3	99.00	192.743	0.0808	
4	75.00	385.485	0.4267	
5	0.00	770.970	> 999 *	
6	86.00	1541.941	0.2442#	
7	56.00	3083.881	0.9107#	
8	27.00	6167.763	2.9630#	
9	13.00	12335.530	7.2308#	
10	3.00	24671.051	34.6667	
11	1.00	49342.102	106.0000*	
12	1.00	98684.203	106.0000*	
CONTROL It'	s: 96.00	105.00 120	00	
10 044444444	4444444444	****	. <b>00</b>	
*SLOPE - 1.	6366			
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100	1000	CONCENTRATION	10E4	10E5

EC50 3441.762 (95% CONFIDENCE RANGE: 2795.067 TO 4238.083)

FILE NAME: 6-2-2-3B.SPT TEST DATE: TEST TIME: Sample Description: Room temp. 0x. 9°C 2:40pm, 2-12-93, Bryrant's Creek, Detailed Test, Dupe B Procedure: SOLID-PHASE Osmotic Adjustment: none Dilution Factor : 2 Concentration Units: ppm Initial Concentration: 98684,2 ppm Test Time: 5 minutes number Ιt CONC. GANNIA 55.00 53.00 57.00 48.186 96.371 192.743 -0.0061\* 1 0.0314\* -0.0409\* 385.485 0.1389 5 49.00 770.970 0.1156# 39.00 26.00 15.00 1541.941 3083.881 6167.763 0.4017# 1.1026# 2.6444# 5.00 12335.530 9.9333# 10 2.00 24671.051 26.3333# 11 12 1.00 49342.102 53.6667 > 999 \* 0.00 98684.203 CONTROL It's : 95.00 16.00 53.00 \*SLOPE - 1.5515 . EC50 10 844444444444 144**~4444444**. **44444444**46**~4444**44444444 100 1000 CONCENTRATION 10E4

BC50 2975.663 (95% CONFIDENCE RANGE: 2729.961 TO 3243.478)

FILE NAME: 3-WINTER.SPT TEST DATE: 3-1-1-8C TEST TIME: Sample Description: Room temp. 22.26 3:20pm, 2-10-93, Keithsburg (Winter), Detailed Test, Dupe A Procedure: SOLID-PHASE Osmotic Adjustment: none Dilution Factor: 2 Initial Concentration : 98684.2 ppm Test Time: 5 minutes Concentration Units: ppm NUMBER Ιt CONC. GAMMA ----1 92.00 48.186 0.0036\* 96.371 192.743 39.00 1.3675# 3 76.00 0.2149# 385.485 770.970 65.00 0.4205# 34.00 21.00 1.7157# 1541.941 3.3968# 9.2593# 9.00 4.00 4.00 3083.881 22.0833# 6167.763 12335.530 22.0833# 10 2.00 24671.051 49342.102

\*SLOPE - 0.8317

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2.00

45.1667#

45.1667#

. EC50 1000 CONCENTRATION 10E4

EC50 436.952 (95% CONFIDENCE RANGE: 201.172 TO 949.073)

# Appendix E Sediment Elutriate Toxicity Test Results

# HYGIENG 2 LABORATORY THE Unrearity of time.

## UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

Keithsburg Division Site Mark Twain NWR	Source Lab Replicate #1
Sample Type sediment Date Collecte Test Organism <u>Pimephales promelas</u> Age	
IC <sub>50</sub> 5 Chemical and Toxi	23
Test Begun on 8/3/92 Test Ended on 8/7/92	at 930 hrs
Initial	48 hours <b>96 hours</b> 3.1 <b>2.0</b>
Total NH <sub>3</sub> (mg/L) 4.6 Unionized NH <sub>3</sub> (mg/L)	0.43

				FATHEAD MINNOW MORTALITY*					
CHEM	ICAL AND		AL		SAMPLE			CONTRO	L
Time	Temp ('C)	DO	Яq	a	b	С	a	b	С
Initial	24.0	5.1	7.7						
24 hours	26.0	8.1	8.3				TO A SHARE OF THE		
48 hours	27.0	8.0	8.3						
72 hours	27.0	8.6	8.1					ļ	
96 hours	25.0	7.5	8.1	0/7	0/7	0/6	0/7	ا مری	0/6
TOTAL	L HORTALI	TY AT S	6 BOURS			0/20			0/20

\* Mortality - Number Dead / Number Tested

Connents:

Analyst: JLS/JCH Date Reported: AUG 1 1 1992 Verified: John

## UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

Site Mark Twain NWR		Source Lab Repl	
Sample Type sediment			
Organism Pimephales prom	IC <sub>50</sub> 5.7		Reference Toxicant NaCI
Che	maical and Tomic	city Test Data	
T 1			
lest	Begun on <u>8/3/92</u>	_ at <u>930 h</u>	re
•	Begun on <u>8/3/92</u> Ended on <u>8/7/92</u>		
•	Ended on <u>8/7/92</u>		
Test 1	Ended on <u>8/7/92</u>	at <u>930</u> h	re .
Test 1	Ended on 8/7/92 Initial	at 930 h	96 hours

					FATHEAD MINNOW MORTALITY™					
CHEMICAL AND PHYSICAL ANALYSIS				SAMPLE			CONTROL			
Time	Temp (°C)	DO	РЩ	a	Ъ	С	a	ь	С	
Initial	24.0	5.0	7.7							
4 bours	26.0	8.0	8.4	_						
48 bours	25.0	-8-1-	_6.3	-						
72 hours	27.0	8.5	8.3							
96 hours	25.0	7.7	8.3	0/7	0/7	0/6	0/7	1/7	0/6	
TOTAL	L MORTALI	TY AT 9	6 BOURS			0/20			1/20	

\* Mortality = Number Dead / Number Tested

Connents:

Analyst: JLS/JGH AUG 1 1 1992

Verified: John

### REGENC ABORATORY The discrete CHARLES

## UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

A SECOND	DML Lab #	9264461		
Site Keithsburg Division M	T-3-5-2-2	Source Lab Re	plicate #1	
Sample Type Sediment D	ate Collecte	d 9/14/92 Date	Received 9/17	/92
Test Organism Pimephales promela	s Age	16 days Re	ierence Toxicant	NaC1
	IC <sub>50</sub> _5	.73 (48 hour)		
Chenic	al and Toxi	city Test Data		
		at 1030 h		
	Initial	10 hours	96 hours	
NH <sub>3</sub> -N (mg/L)			2.6	
Total NH <sub>3</sub> (mg/L)	10.6			
Unionized NH <sub>3</sub> (mg/L)	0.27	1.0	.0.31	
		FATHEAD MINNO	HORTALITY*	
CHEMICAL AND PHYSICAL ANALYSIS	5	SAMPLE	CONTROL	

				FATHEAD MINNOW MORTALITY*					
CHEMICAL AND PHYSICAL ANALYSIS			SAMPLE			CONTROL			
Time	Temp (°C)	DO	Hq	a	ь	С	a	ь	С
Initial	23.0	5.5	7.7						
24 hours	24.0	7.9	8.4						
48 hours	24.0	8.6	8.4						
hours	24.0	7.2	8.3						
96 hours	25.0	7.3	8.3	3/7	1/7	0/6	0/7	0/7	0/6
TOTA	L MORTALI	<b>TY AT</b> 9	6 BOURS			4/20			0/20

\* Mortality \* Number Dead  $\times$  Number Tested

Comments:

Analyst: JLS/JCM Date Reported: Verified:



#### UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

#### DML Lab # 9264461

	Site_Ke	ithsburg D	ivision l	1T-3-5-2-2	_ Source	<u>مما</u>	Replicate #	2	
	Sample	Type Sed	iment D	ate Coll	ected_9/1	4/92	Date Recei	ved_ 9/17/92	
Test	Organisa	Pimepha	les prome	Las	Age16	days	Reference	Toxicant_NaC	:1
				IC <sub>50</sub>	5,73	(48 h	our)		
			Chenic	al and 1	Coxicity	Test I	Data		

Test Begun on 9/21/92 at 1030 hrs Test Ended on 9/25/92 at 1030 hrs

		Initial	48 hours	96 hours
ин <sup>3</sup> -и	(mg/L)	9.2	7.2	2.8
Total NH3	(mg/L)	11.2	8.7	3.4
Unionized NH	(mg/L)	0.22	.0:	0.35

					FATHE	AD MINNO	W MORT	ALITY*	
CHER	ICAL AND ANALYS		AL		SAMPLE			CONTRO	L
Time	Temp (°C)	DO	Hq	ā	ь	С	a	b	С
Initial	23.0	5.5	7.6						
24 hours	24.0	7.9	8.4						
48 hours	24.0	8.3	8.4						
72 hours	24.0	7.2	8.4						
96 hours	25.0	7.1	8.3	1/7	2/7	1/6	0/7	0/7	0/6
TOTAL	L MORTALI	TY AT 9	6 BOURS			4/20			0/20

\* Mortality \* Number Dead / Number Tested

Comments:

Analyst: JLS/JGM Date Reported:



## UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

De produ			.1	DML Lab #	926446	2			
Site	B_ Keiths	burg Div	ision MI	-3-5-3-2	Source	Lab Re	plicate i	1	
Sax	aple Type	Sedime	nt Da	te Collec	ted_9/1	4/92 Da	te Rece:	ived_9/1	7/92
est Organ	nisa <u>Pi</u>	mephales	promela	s A	ge <u>16</u>	days R	eference	Toxica	nt <u>NaC</u>
						(48 hour)			
			Chenica	al and To	<b>x</b> icity	Test Dat	a		
		т							
				n on <u>9/2</u> n on <u>9/2</u>					
		WU V	/ /T \	Initial					
		•		7.8				.1	•
	Tot	al NH <sub>3</sub>	(mg/L)	9.5		7,8	3	1	•
	Unioniz	ed NH <sub>3</sub>	(mg/L)_	0.18		0.92	0	.34	
					FATH	EAD MINN	OW MORT	ALITY*	
CHEM	ICAL AND		AL		SAMPLE			CONTROL	
Time	Temp (°C)	DO	Hq	a	b	С	a	b	С
Initial	23.0	5.6	7.6						
4 hours	24.0	8.0	8.4						
B hours	24.0	8.4	8.4						
2 hours	24.0	7.1	8.2	200000000000000000000000000000000000000					
6 hours	25.0	7.2	8.3	0/7	2/7	2/6	.0/7	0/7	0/6
2021	L MORTALI				·	4/20			0/20
IUIA									

Comments:

Analyst: JLS/JGM Date Reported: Verified: H



#### UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

`	40001051		DHL Lab #	9264462		
	Site_K	ithsburg Division 1	rr-3-5-3-2	Source Lab Repl	icate #2	
	Sample	Type Sediment D	ate Collecte	sd_9/14/92 Date	Received 9/17/92	
Test	Organism	Pimephales promel	.as Age	e <u>16 days</u> Ref	erence Toxicant NaC	1
			IC <sub>50-5</sub>	.73 (48 hour)		
		Cheaic	al and Toxi	icity Test Data		
		Test Begu	un con <u>9/21/9</u>	92 at 1030 h	re	
		Test Ende	nd on 9/25/9	02 at 1030 h	rs	
			Initial	48 hours	96 hours	
		NH3-N (mg/l)	8.3	6.6	2.2	
		Total NH <sub>3</sub> (mg/L)	10,1	8.0	2.7	
	Un	ionized NH <sub>3</sub> (mg/L)	0.19	1.2	0.34	

					FATHE	AD MINN	OW MORT	ALITY*	
CHE	ICAL AND		AL		SAMPLE			CONTRO	ī.
Time	Temp (°C)	DO	Hq	a	ь	С	a	ь	С
Initial	23.0	5.6	7.6						
24 hours	24.0	8.3	8.4						
48 hours	24.0	8.4	8.5						
72 hours	24.0	7.6	8.4						
96 hours	25.0	7.7	8.4	1/7	0/7	0/6	0/7	0/7	0/6
TOTA	L MORTAL	TY AT 9	6 HOURS			1/20			0/20

\* Mortality \* Number Dead / Number Tested

Comments:

Analyst: JLS/JGM Date Reported:

Verified:



## UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

#### DML Lab #

Site Control S	ource Lab Replicate # 1
Sample Type water Date Collected	Date Received
Tout Organism <u>Pimephales promelas</u> Agu	9 days Reference Toxicant NaCl
IC <sub>50</sub> 5.7	3
Chemical and Toxic	ity Test Data
Test Begun on <u>8/3/92</u>	at 930 hrs
Test Ended on 8/7/92	at 930 hrs
· Initial	48 hours 96 hours
NH <sub>3</sub> -N (mg/L) <u>&lt;0.1</u>	0.2 0.2
Total NH <sub>3</sub> (mg/L) <0.1	0.2
Unionized NH <sub>3</sub> (ag/L) <0.01	0.02 0.02

					FATHE	PD RINN	OW MORT	ALITY*	
CHEN	ICAL AND		I		SAMPLE			CONTRO	L
Time	Temp (°C)	DO	Ηq	a	b	С	a	b	C
Initial	24.0	11.2	8.3						
24 hours	26.0	8.3	8.3						
48 hours	26.0	8.4	8.3						
/2 hours	27.0	8.8	8.2						
96 hours	25.0	8.2	8.3				0/7	0/7	0/6
TOTA	L MORTAL	TTV 1T 0	( FOTTE						0/20

\* Mortality = Number Dead / Number Tested

Comments:

Analyst: JLS/JCM Date Reported: AUG 1 1 1992 Verified:

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## UNIVERSITY HYGIENIC LABORATORY TOXICITY TEST

#### DML Lab #

	c Cont	rol			Source	Lab R	eplicate	1 2	
Sa	mple Type	water	Dat	e Collec	ted_	Da	te Recei	i ved	
Test Orga	nism <u>Pime</u>	phales p	romelas	A	ge 9 day	<u>s</u> R	eference	Toxican	t_NaCl
				IC <sub>50</sub> —	5.73				
		(	Chenica.	l and To	zicity T	est Dat	<b>a</b>		
		Tes	t Begun	on 8/3/	92 at	930	hve		
					92 at _				
			-	Initial	48	hours	96	hours	
		ин <sup>3</sup> -и (	mg/L) _	<0.1		0.1		0.3	
	Tot	al NH <sub>3</sub> (	mg/L)	<u>&lt;0</u>		0.1	ک ،	1.4	
	Unioniz	ed NH <sub>2</sub> (	, mg/L) _	<0.01		0.01		0.04	
		,							
					FATHE	AD MINN	OW MORT	ALITY*	
CHE	DILA LASIN ANALYS		L		SAMPLE			CONTROL	
Time	Temp (°C)	DO	Нq	a	b	С	a	þ	С

CHE	ICAL AND ANALYS		ΔI.		SAMPLE			CONTRO	
Time	Temp (°C)	DO	рĦ	a	ь	С	a	b	С
Initial	24.0	11.0	8.3						
24 hours	26.0	8.4	8.3						
48 hours	27.0	8.5	8.2						
72 hours	25.0	8.9	8.2						
96 hours	25.0	8.2	8.3				0/7	1/7	0/6
TOTA	L MORTALI	TY AT	6 HOURS						1/20

\* Mortality \* Number Dead / Number Tested

Comments:

Analyst: JLS/JGM AUG 1 1 1992

Verified

# Appendix F Water Quality Data Records

Appendix F. Water quality data for the sampling locations at Mark Twain National Widlife Refuge study area, 1992.

BITE	REPERENCE NUMBER	DATE	TIME	WATER DEPTH	WATER TEMP	CONDUCTIVITY	DO-top	DO-botto
				(meters)	(celcius)	(uS/cm)	(mg/L)	(mg/L)
Strunk Steuch	1	9-Jun-92	1200	1.00	27.00	440.00	6.20	
and an entire of the second of	ż	9-Jun-92	1300	2.50	27.00	470.00	8.00	
	3	9-Jun-92	1500	2.50	26.00	440.00	10.00	
Mg Timber Division	•	22-May-92	1030	6.50	23.00	310.00	7.50	
	ì	22-May-92	1130	2.00	25.00	320.00	11.00	
	i	22-May-92	1300	4.00	20.00	•		
	7	22-May-92	1400	3.00			•	
	à	22-May-92	1500	3.00				
	ř	11-Jun-92	930	0.50	23.50	400.00	8.30	
	10	11-Jun-92	1100	0.90	25.00	400.00	11.00	
	11	11-Jun-92	1300	3.50	22.00	390.00	11.20	5.80
	12	11-Jun-92	1500	4.00	23.00	370.00	14.00	6.20
Ceithsburg Division	13	19-May-92	1030	5.00	21.20	340.00	5.20	3.50
CALCIDOCA CAMPON	14	19-May-92	1130	5.50	21.20	340.00	8.30	3.40
	15	19-May-92	1300	2.78	20.50	340.00	15.00	10.20
	16	19-May-92	1400	2.75	20.20	310.00	15.00	11.30
	17	19-May-92	1630	5.25	22.50		13.00	12.00
	18	14-Jul-92	1000	3.50	25.50	280.00	4.10	0.50
	19	14-Jul-92	1200	4.00	28.00	315.00	4.40	3.60
	13/14/17a	14-Sep-92	1000	2.50	19.00	430.00	2.00	1.00
	13/14/17b	14-Sep-92	1030	2.50	19.00	455.00	2.20	0.00
	13/14/17c	14-Sep-92	1100	2.60	20.00	470.00	2.00	1.20
	13/14/176	14-840-92	1130	2.50	20.00	350.00	4.20	3.80
	13/14/170	14-Sep-92	1200	2.50	20.00	350.00	5.70	4.00
ardner Division	20	18-May-92	1000	1.75	19.50	310.00	8.10	
	21	18-May-92	1100	1.75				
	22	18-May-92	1130	1.50	20.50	355.00	8.30	
	23	18-May-92	1200	1.75	21.50	465.00	9.20	
Clarence Cannon Refug		14-May-92		1.75				
	25	14-May-92		2.00	19.00	190.00	10.50	
	26	14-May-92		3.00				
	27	14-May-92		3.00	19.00	200.00	11.00	
	28	14-May-92		4.00	18.00	430.00	7.50	
	29	14-May-92		3.50				
	30	14-May-92		5.60	17.00	290.00	12.00	
	31	14-May-92		11.00	17.00	280.00	12.00	
Letchtown Division	32	12-May-92	1000	4.00	21.00	285.00	13.80	
	33	12-May-92	1015	4.00				
	34	12-May-92	1035	4.50	21.50	295.00	11.70	9.90
	35	12-May-92	1045	4.50	21.50	295.00	15.00	13.40
	36	12-May-92	1130	4.50				
	37	12-May-92	1200	4.00	21.00	319.00	14.60	
	38	12-May-92	1300	4.50	23.00	310.00	15.00	15.00
	39	12-May-92	1330	2.50				
	40	13-May-92	1200	2.50	20.50	250.00	9.70	_
	41	13-May-92	1230	3.00	21.00	310.00	13.50	-
	42	13-May-92	1245	3.50	21.00	350.00	13.00	
	43	13-Mey-92	1410	4.00	17.50	335.00	5.40	1.00

## Appendix G

**Quality Assurance and Quality Control Test Results** 

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

#### PROCEDURAL BLANKS

Analyte	Lab Sample Num	ber Result Total UG
% Moisture	BLANK-B	
	BLANK-F	
	BLANK-G	
	BLANK-J	
	BLANK-N	
	BLANK-P	
Al	BLANK-B	-47
	BLANK-F	4
	BLANK-G	4
	BLANK-J	26
	BLANK-N	8
	BLANK-P	4
As	BLANK-B	02
	BLANK-F	.01
	BLANK-G	02
	BLANK-J	01
	BLANK-N	.03
	BLANK-P	01
Cd	BLANK-B	0
	BLANK-F	0
	BLANK-G	0
	BLANK-J	0
	BLANK-N	<b>O</b> :
	BLANK-P	0
Cr	BLANK-B	5
	BLANK-F	.04
	BLANK-G	-1.09
	BLANK-J	1.5
	BLANK-N	.01
	BLANK-P	.03

Catalog: 3070021 Lab Name: GERG 30-Jun-93 Purchase Order: 85830-2-3836

Page: 48

### PROCEDURAL BLANKS (Cont.)

Analyte	Lab Sample Nu	Result Total UG	
Pb	BLANK-B	.07	
	BLANK-F	.02	
	BLANK-G		
	···· <del>-</del>	.02	
	BLANK-J	0	
	BLANK-N	.03	
	BLANK-P	.02	
Se	BLANK-B	.01	
	BLANK-F	0	
	BLANK-G	02	
	BLANK-J	.03	
	BLANK-N	0	
	BLANK-P	Ō	
Zn	BLANK-B	9	
	BLANK-F	-8.1	
	BLANK-G	-7.9	
	BLANK-J	7.4	
	BLANK-N	4.7	
	BLANK-P	4.8	

Catalog: 3070021

Page: 49

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

#### **DUPLICATES**

A	Sample	ample Initial Result Duplicate Result		Relatio	Relative %		
Analyte	Numb	er Sample Matrix	(ppm / %)	(ppm / %)	Averag		
% Moisture		1-1 Sediments	38.9 %	43.3 %	 41.	1 10.71	
	3-2-1-1	Sediments	52.8 %	53.4 %	53.1	1.13	
	4-1-3-2	Sediments	36.9 %	37.3 %	37.1	1.13	
	5-1-3-1	Sediments	51.2 %	46.1 %	48.65	1.08	
	6-1-4-1	Sediments	49.9 %	49.3 %	49.6		
	7-1-3-1	Sediments	31 %	30.7 %	30.85	1.21 0.97	
AVS	0-1-1-1	Sediments	564.7 Dry	603 Dry	583.85	6.56	
	3-2-1-1	Sediments	377 Dry	391.8 Dry	384.4	0.00	
	4-1-3-2	Sediments	174.7 Dry	155.4 Dry		3.85	
	5-1-3-1	Sediments	805.9 Dry	743.3 Dry		11.69	
	6-1-4-1	Sediments	103.4 Dry	99.3 Dry		8.08	
	7-1-3-1	Sediments	27.5 Dry	28.1 Dry	101.35 27.8	4.05 2.16	
Al		Sediments	19917 Dry	23788 Dry	21852.8	5 17.71	
		Sediments	12626 Dry	11696 Dry	12161	7.65	
		Sediments	7967 Dry	11578 Dry	9772.5		
		Sediments	22142 Dry	21039 Dry	21590.5		
		Sediments	8046 Dry	10306 Dry	9176		
	7-1-3-1	Sediments	7261 Dry	10695 Dry	8978	24.63 38.25	
As		Sediments	5.84 Dry	7.43 Dry	6.635	22.00	
		Sediments	6.54 Dry	7.05 Dry	6.795	23.96	
		Sediments	4.24 Dry	4.46 Dry	4.35	7.51	
Į.	5-1-3-1	Sediments	5.64 Dry	5.48 Dry	5.56	5.06	
(	6-1-4-1	Sediments	3.06 Dry	3.05 Dry		2.88	
7	7-1-3-1	Sediments	5.3 Dry	5.63 Dry	3.055 5.465	0.33 6.04	
Cd	0-1-1-1	Sediments	.38 Dry	.42 Dry	0.4	40	
3	3-2-1-1	Sediments	.58 Dry	.63 Dry	0.4 0.605	10	
4	I-1-3-2 S	Sediments	.41 Dry	.41 Dry		8.26	
		Sediments	.52 Dry	.56 Dry	0.41	0	
•	3-1-4-1 8	Sediments	.35 Dry	.38 Dry		7.41	
7	7-1-3-1 8	Sediments	.33 Dry	.35 Dry		8.22 5.88	

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

### DUPLICATES (Cont.

Relative %	Relative %		
	ifference		
 Dry 21,105 2	20.8		
	20.8 ).72		
	9.72 B.73		
_ • • • • • • • • • • • • • • • • • • •	6.73 .13		
Ory 8.995 24.			
Ory 11.2 27.6			
Dry 28.665 1	18.18		
	.04		
	.04 94		
	.71		
Dry 22.57 5,			
Ory 14.21 5.3			
Dry 27482	10.20		
	19.39 5.01		
	1.23		
	1.23		
	9.87		
_ ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' ' '	9.67  .43		
Ory 0.05 o			
	,		
y 0.05 <b>0</b>			
y 0.05 <b>0</b>			
y 0.05 <b>0</b>			
y 0.05 <b>o</b>			
Ory 776.5 22:			
	-		
	7 355.5 9.28 7 458.5 8.94		

Catalog: 3070021

Page: 51

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

#### **DUPLICATES (Cont**

	Sample	Initial Result Duplicate Result			Relative %		
Analyte	Numb	er Sample Matrix	(ppm / %)	(ppm / %)	Average	Difference	
Ni	0-1-1-1	Sediments	20.9 Dry	25.3 Dry	23.1	19.05	
	3-2-1-1	Sediments	20.6 Dry	18.1 Dry	19.35	12.92	
	4-1-3-2	Sediments	11.3 Dry	12 Dry	11.65	6.01	
	5-1-3-1	Sediments	29 Dry	29 Dry	29	0	
	6-1-4-1	Sediments	13 Dry	15.1 Dry	14.05	14.95	
	7-1-3-1	Sediments	17.5 Dry	19.5 Dry	18.5	10.81	
Рь	0-1-1-1	Sediments	51.92 Dry	45.33 Dry	48.625	13.55	
	3-2-1-1	Sediments	17.96 Dry	16.72 Dry	17.34	7.15	
	4-1-3-2	Sediments	13.15 Dry	11.82 Dry	12.485	10.65	
	5-1-3-1	Sediments	20.74 Dry	21.95 Dry	21.345	5.67	
	6-1-4-1	Sediments	17.03 Dry	17.77 Dry	17.4	4.25	
	7-1-3-1	Sediments	17.96 Dry	19.54 Dry	18.75	8.43	
Se	0-1-1-1	Sediments	< 1 Dry	< 1 Dry	0.5	0	
	3-2-1-1	Sediments	< 1 Dry	< 1 Dry	0.5	0	
	4-1-3-2	Sediments	< 1 Dry	< 1 Dry	0.5	0	
	5-1-3-1	Sediments	< 1 Dry	< 1 Dry	0.5	0	
	6-1-4-1	Sediments	< 1 Dry	< 1 Dry	0.5	0	
	7-1-3-1	Sediments	< 1 Dry	< 1 Dry	0.5	0	
Zn	0-1-1-1	Sediments	88.6 Dry	109.8 Dry	99.2	21.37	
	3-2-1-1	Sediments	76.2 Dry	80.1 Dry	78.15	4.99	
	4-1-3-2	Sediments	56.4 Dry	68.9 Dry	<b>57.65</b>	4.34	
	<b>5-1-3-1</b>	Sediments	103.9 Dry	105.6 Dry	104.75	1.62	
	6-1-4-1	Sediments	55.3 Dry	65.6 Dry	60.45	17.04	
	7-1-3-1	Sediments	50.7 Dry	58.3 Dry	54.5	13.94	

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

#### REFERENCE MATERIALS

	Lab Sam	-1-		* Certified	95%			
Δna	Lab Samı İyte Numi			Refere		Result	Percent	
	overy	ber S.R.M. ID	S.R.M	. Name	Value (ppm / %)	Interval	(ppm / %)	
Al	BCSS-D	NRCC BCSS-1	<b>*****</b>				40.5	
	BCSS-E	NRCC BCSS-1					19 Dry	
	BCSS-I						8 Dry	
	BCSS-L	NRCC BCSS-1				21829	•	
	BCSS-M	NRCC BCSS-1					0 Dry 91 Dry	
As	BCSS-D		Sediment		11.1 Dry	1.4	9 61 Dm.	06.50
	BCSS-E	NRCC BCSS-1	Sediment		11.1 Dry	1.4	9.61 Dry 9.58 Dry	86.58
	BCSS-I	NRCC BCSS-1	Sediment		11.1 Dry	1.4	8.4 Dry	86.31 75.68
	BCSS-L		Sediment		11.1 Dry	1.4	8.19 Dry	78.78
	BCSS-M	NRCC BCSS-1	Sediment		11.1 Dry	1.4	9.65 Dry	
Cd	BCSS-D		Sediment		.25 Dry	.4	.25 Drv	100
	BCSS-E	NRCC BCSS-1	Sediment		.25 Dry	.4	.25 Dry	100
	BCSS-I	NRCC BCSS-1	Sediment		.25 Dry	.4	.23 Dry	92
	BCSS-L	NRCC BCSS-1	Sediment		.25 Dry	.4	.24 Dry	96
	BCSS-M	NRCC BCSS-1	Sediment		.25 Dry	.4	.23 Dry	92
Cr	BCSS-D	NRCC BCSS-1	Sediment		123 Dry	14	40.95 Dry	33.29
	BCSS-E	NRCC BCSS-1	Sediment		123 Dry	14	51.13 Dry	41.57
		NRCC BCSS-1	Sediment		123 Dry	14	49.91 Dry	40.58
	BCSS-L	NRCC BCSS-1	Sediment		123 Dry	14	53.12 Dry	43.19
	BCSS-M	NRCC BCSS-1	Sediment		123 Dry	14	50.42 Dry	40.99
Cu	BCSS-D	NRCC BCSS-1	Sediment		18.5 Dry	2.7	14.33 Dry	77.46
		NRCC BCSS-1	Sediment		18.5 Dry	2.7	18.52 Dry	100.11
		NRCC BCSS-1	Sediment		18.5 Dry	2.7	18.76 Dry	101.41
		NRCC BCSS-1	Sediment		18.5 Dry	2.7	18.57 Dry	100.38
	BCSS-M	NRCC BCSS-1	Sediment		18.5 Dry	2.7	18.44 Dry	99.68
Fe	BCSS-D	NRCC BCSS-1				2309	2 Dry	

<sup>\*</sup> Only certified analytes list a confidence interval - all others are considered reference values.

Catalog: 3070021 Lab Name: GERG 30-Jun-93 Purchase Order: 85830-2-3836

Page: 53

### REFERENCE MATERIALS (Cont.

	Lab Sam			* Certified	95%			
Δns	alyte Numi			Refere	nce Confidenc	e Resul	t Percen	nt
Rec	overy	ber S.R.M. ID	S.R.M.	Name	Value (ppm / %)	interval	(ppm / %)	
Fe	BCSS-E	NRCC BCSS-1				210	004 D= :	
	BCSS-I	NRCC BCSS-1					94 Dry	
	BCSS-L	NRCC BCSS-1					2 Dry	
	BCSS-M	NRCC BCSS-1					27 Dry 68 Dry	
Hg	BEST-A	NRCC BEST-1	Sediment		.092 Dry	.009	, 1 D	
	BEST-D	NRCC BEST-1	Sediment		.092 Dry	.009	< .1 Dry	
	BEST-E	NRCC BEST-1	Sediment		.092 Dry	.009	< .1 Dry	108.7
	BEST-I	NRCC BEST-1	Sediment		.092 Dry	.009	< .1 Dry	108.7
	BEST-L	NRCC BEST-1	Sediment		.092 Dry	.009	< .1 Dry < .1 Dry	108.7
	BEST-M	NRCC BEST-1	Sediment		.092 Dry	.009	< .1 Dry	108.7 108.7
Mn	BCSS-E	NRCC BCSS-1	Sediment		229 Dry	15		
	BCSS-E	NRCC BCSS-1	Sediment		229 Dry	15	180 Dry	
	BCSS-I	NRCC BCSS-1	Sediment		229 Dry	15	228 Dry	99.56
	BCSS-L	NRCC BCSS-1	Sediment		229 Dry	15	231 Dry	100.87
	BCSS-M	NRCC BCSS-1	Sediment		229 Dry	15	229 Dry 219 Dry	100 95.63
Ni	BCSS-D	NRCC BCSS-1	Sediment		55.3 Dry	2.0	40.00	
	BCSS-E	NRCC BCSS-1	Sediment		55.3 Dry	3.6	46.6 Dry	84.27
	BCSS-I	NRCC BCSS-1	Sediment		55.3 Dry	3.6	56.03 Dry	101.32
	BCSS-L	NRCC BCSS-1	Sediment		55.3 Dry	3.6 3.6	60.21 Dry	108.88
	BCSS-M	NRCC BCSS-1	Sediment		55.3 Dry	3.6	58.86 Dry	106.44
					00.5 Liy	3.0	57.7 Dry	104.34
Pb	BCSS-D	NRCC BCSS-1	Sediment		22.7 Dry	3.4	21.04.0	00.05
	BCSS-E	NRCC BCSS-1	Sediment		22.7 Dry	3.4	21.94 Dry 21.99 Dry	96.65
		NRCC BCSS-1	Sediment		22.7 Dry	3.4	20.55 Dry	96.87
	BCSS-L	NRCC BCSS-1	Sediment		22.7 Dry	3.4	21.35 Dry	90.53
	BCSS-M	NRCC BCSS-1	Sediment		22.7 Dry	3.4	21.05 Dry	94.05 92.73
Se Se	BCSS-D	NRCC BCSS-1	Sediment		.43 Dry	.06		
	BCSS-E	NRCC BCSS-1	Sediment		.43 Dry		< 1 Dry	232.56
		<del> •</del>			.43 Dry	.06	< 1 Dry	232.56

<sup>\*</sup> Only certified analytes list a confidence interval - all others are considered reference values.

Catalog: 3070021 Lab Name: GERG

Page: 54

30-Jun-93

Purchase Order: 85830-2-3836

#### REFERENCE MATERIALS (Cont.)

	Lab Sample		* Cer	tified 95 Reference	% Confidence	Result	Percent	
Anal Reco	lyte Number overy	S.R.M. ID	S.R.M. Nam	ne Value	(ppm / %)	Interval	(ppm / %)	
Se	BCSS-L NRC	CC BCSS-1 CC BCSS-1 CC BCSS-1	Sediment Sediment Sediment		.43 Dry 43 Dry .43 Dry	.06 .06 .06	< 1 Dry < 1 Dry < 1 Dry	232.56 232.56 232.56
Zn	BCSS-E NRC BCSS-I NRC BCSS-L NRC	CC BCSS-1 C BCSS-1 C BCSS-1 C BCSS-1 CC BCSS-1	Sediment Sediment Sediment Sediment Sediment	1 1 <sup>1</sup> 1	119 Dry 19 Dry 19 Dry 19 Dry 119 Dry	12 12 12 12 12	102 Dry 116.6 Dry 120.5 Dry 121.8 Dry 115.3 Dry	85.71 97.98 101.26 102.35 96.89

<sup>•</sup> Only certified analytes list a confidence interval - all others are considered reference values.

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

#### **SPIKE RECOVERIES**

	Sample		Spike Level Ame	ount Recovered	* Spike /	Percent
Analyte	Numb			(ppm / %)	Backgro	und Recovery
AVS	0-1-1-2		893.3 Dry			77.67
	3-2-1-2	Sediments	250.6 Dry			93.14
	4-2-1-1	Sediments	376.1 Dry	345.8 Dry		
	5-1-3-2	Sediments	664.1 Dry	611 Dry		92
	6-1-4-2	Sediments	436.7 Dry	407 Dry		93.2
	7-1-3-2	Sediments	226.1 Dry	206.4 Dry		91.29
As	0-1-1-2	Sediments	7 Dry	7.13 Dry	1.07	101.86
	3-2-1-2	Sediments	7 Dry	5.45 Dry	1.05	77.86
	4-2-1-1	Sediments	6 Dry	6.18 Dry	0.73	103
	5-1-3-2	Sediments	7 Dry	6.21 Dry		88.71
	6-1-4-2	Sediments	7 Dry	7.91 Dry		113
	7-1-3-2	Sediments	7 Dry	6.34 Dry		90.67
Cd	0-1-1-2	Sediments	10 Dry	9.93 Dry	23.26	99.3
	3-2-1-2	Sediments	9 Dry	11 Dry		122.22
	4-2-1-1	Sediments	9 Dry		15.25	115.89
	5-1-3-2	Sediments	10 Dry		19.23	99.4
	6-1-4-2	Sediments	10 Dry	10.24 Dry		102.4
	7-1-3-2	Sediments	10 Dry	10.65 Dry		106.5
<b>Cr</b>	0-1-1-2	Sediments	53 Dry	61.13 Dry	3.15	115.34
	3-2-1-2	Sediments	52 Dry	50.21 Dry		96.56
	4-2-1-1	Sediments	51 Dry	41.61 Dry		81.59
	5-1-3-2	Sediments	53 Dry	53.23 Dry		100.43
	6-1-4-2	Sediments	65 Dry	72.53 Dry		131.87
	7-1-3-2	Sediments	55 Dry	65.33 Dry	2.73	118.78
Cu	0-1-1-2	Sediments	56 Dry	53.48 Dry	1.84	95.5
	3-2-1-2	Sediments	64 Dry	52.08 Dry	2.5	96.44
	4-2-1-1	Sediments	38 Dry	41.65 Dry		109.61
	5-1-3-2	Sediments	55 Dry	49.5 Dry		90

<sup>\*</sup> For a spike to be a valid measure of method accuracy, this ratio must be higher than 1.0.

Catalog: 3070021 Lab Name: GERG 30-Jun-93 Purchase Order: 85830-2-3838

Page: 56

#### **SPIKE RECOVERIES (Cont.)**

	Sample		Spike Level	Α	mount R	ecov	ered	• Spike /	Per	ent
Analyte		er Sample Matri:		n / %)			n / %)	Backg	round	Recovery
Cu	6-1-4-2			Dry				2.5	10	3.63
	7-1-3-2		67	Dry			-	2.6	104	.88
Hg	0-1-1-2	Sediments	1.27	7 Dry		1.29	Dry	12.7		01.57
	3-2-1-2	Sediments	1.27	Dry	•	1.36	Dry	12.7	10	7.09
	4-2-1-1	Sediments	1.27	Dry	•	1.36	Dry	12.7	10	7.09
	<b>5-1-3-2</b>	Sediments	1.27	Dry	•	1.32	Dry	12.7	10	3.94
	6-1-4-2	Sediments	1.27	Dry	•	1.33	Dry	12.7	10	4.72
	7-1-3-2	Sediments	1.27	Dry	•	1.28	Dry	12.7	10	D. <b>79</b>
Mn	0-1-1-2	Sediments	81:	3 Dry		86	5 Dry	0.94	1	06.4
	3-2-1-2	Sediments	659	Dry		870	Dry	0.58	13:	2.02
	4-2-1-1	Sediments	370	Dry		440	Dry	0.69	118	3.92
	5-1-3-2	Sediments	671	Dry		743	Dry	0.59	110	0.73
	6-1-4-2	Sediments	697	Dry		755	Dry	2	108.	32
	7-1-3-2	Sediments	694	Dry		803	Dry	1.11	111	5.71
Ni	0-1-1-2	Sediments	167	Dry		209	Dry	7.91	12	5.15
	3-2-1-2	Sediments	162	Dry			Dry	10.06	10	6.54
	4-2-1-1	Sediments	160	Dry	1	38.4	Dry	9.58	8	6.5
	<b>5-1-3-2</b>	Sediments	165	Dry	1	69.3	Dry	5.34	10	2.61
	6-1-4-2	Sediments	172	Dry	1	84.4	Dry	18.11	10	7.21
	7-1-3-2	Sediments	171	Dry	1	84.9	Dry	6.5	108	3.13
Pb	0-1-1-2	Sediments	248	Dry	3	94.7	2 Dry	9.87	1	59.16
	3-2-1-2	Sediments	241	Dry	24	5.24	Dry	12.49	1	01.76
	4-2-1-1	Sediments	237	Dry	2	<b>55.9</b>	Dry	12.67	10	7. <b>97</b>
	5-1-3-2	Sediments		Dry			Dry			95.5
	6-1-4-2	Sediments	255	Dry	27	2.56	Dry	14.03	1	06.89
	7-1-3-2	Sediments	254	Dry	25	3.74	Dry	10.51		99.9
Se	0-1-1-2	Sediments	46	Dry	6	1.78	Dry	46	11	2.57
	3-2-1-2	Sediments	45	Drv	42	2.51	Drv	45	94.	47

<sup>\*</sup> For a spike to be a valid measure of method accuracy, this ratio must be higher than 1.0

Catalog: 3070021

Page: 57

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

#### SPIKE RECOVERIES (Cont.)

Ana	Sample lyte Numb	er Sample Matr		ount Recovered (ppm / %)	* Spike / Backgr	Percent ound Recovery
Se	4-2-1-1 5-1-3-2 6-1-4-2 7-1-3-2	Sediments Sediments Sediments Sediments	44 Dry 46 Dry 47 Dry 47 Dry	46.56 Dry 43.21 Dry 46.61 Dry 55.87 Dry	44 46 47 47	105.82 93.93 99.17 118.87
Zn	0-1-1-2 3-2-1-2 4-2-1-1 5-1-3-2 6-1-4-2 7-1-3-2	Sediments Sediments Sediments Sediments Sediments Sediments	283 Dry 273 Dry 268 Dry 278 Dry 289 Dry 288 Dry	377.5 Dry 251.9 Dry 218.1 Dry 260.2 Dry 308.8 Dry 311.5 Dry	2.97 3.7 4.08 2.2 5.19 3.63	133.39 92.27 81.38 93.6 106.85 108.16

<sup>\*</sup> For a spike to be a valid measure of method accuracy, this ratio must be higher than 1.0.

COMMENTS (RESULT MODIFERS AND QA/QC COMMENTS) Sample Analyte Number Result Modifier ..... NO DATA EXIST FOR THIS SECTION. QA/QC Comments APPROVAL,EC

Catalog: 3070021 Lab Name: GERG 30-Jun-93 Purchase Order: 85830-2-3836

Page: 58

Catalog: 3070021 Lab Name: GERG 30-Jun-93 Purchase Order: 85830-2-3836

Page: 59

#### **ANALYTICAL METHODS**

Method Code Method Description 002 LABORATORY: Geochemical & Environmental Research Group, Texas A&M

#### Tissue, Sediment and Water Mercury

II. Mercury was determined by EPA method 245.5 with minor revisions. Sediment samples can be analyzed either freeze dried or on a wet basis. Sediment samples are homogenized by mixing before subsampling. The tissue samples were homogenized in the original sample containers with a Tekar Tissumizer and subsampled. Water samples are acidified (0.5% v/v with high purity nitric acid, HNO3) in the original sample bottle. For sediments a 0.5 to 1.0 gram sample (dry weight) was used. For tissues a 1.5 to 2.0 gram sample (wet weight) was used. For water the sample size is 20 ml.

For tissue and sediment, the sample is weighed into a 50 ml polypropylene centrifuge tube. 2.5 ml of concentrated sulfuric acid (H2SO4) and 1.5 ml of concentrated nitric acid (HNO3) were added and the samples heated in a water bath at 90 C for 15 min. After cooling 10 ml of distilled water and 15 ml of mixture of 3.3% (w/w) potassium permanganate (KMnO4), and 1.7% (w/w) potassium persulfate (K2S2O8) were added to each tube and the samples heated in a water bath at 90 C for 30 min. After cooling 5 ml of 10% (w/w) hydroxylamine hydrochloride (NH2OH HC1) was added to reduce excess permanganate and the volume brought to 35 ml with distilled water.

For water samples, the sample is weighed into a 50 ml polypropylene centrifuge tube, 1 ml of concentrated H2SO4 is added and the solution mixed vigorously with a vortex stirrer. Then 4.5 ml of the KMnOr/K2S2O8 is added and the resulting mixture heated in a 90 C water bath for 2 hours. After cooling, 1.5 ml of a 10% (w/w) hydroxylamine hydrochloride (NH2OH HC1) solution is added, sample volume adjusted to a constant volume with distilled water and the resulting solution mixed vigorously.

Lab Name: GERG

30-Jun-93

Purchase Order: 85830-2-3836

#### ANALYTICAL METHODS (Cont.)

Method Code

**Method Description** 

002

Mercury is determined by a modification of the method of Hatch and Ott (1968). A portion of the digest solution is placed in a sealed container. To this is added 0.4 ml of 10% (w/w) stannous chloride (SnCl2). Mercury is reduced to the elemental state and aerated from solution into an atomic absorption spectrophotometer where its concentration is measured.

#### References

- "USEPA Contract Laboratory Program Statement of Work for Inorganic Analysis." Document Number ILMO1.0 USEPA Environmental Monitoring and Support Laboratory, Cincinnati, Ohio.
- "Interim Method for the Sampling and Analysis of Priority
  Pollutants in Sediments and Fish Tissue," USEPA Environmental
  Monitoring and Support Laboratory, Cincinnati, Ohio, Aug. 1977,
  Revised Oct. 1980.
- Hatch, W.R. and Ott, W.L., "Determination of Sub-Microgram Quantities of Mercury in Solution by a Flameles Atomic Absorption Technique", Analytical Chemistry 40,14 p 2085-2087 (1968).

003 LABORATORY: Geochemical & Environmental Research Group, Texas A&M

#### % Dry Weight

III. Approximately 1 gram of wet sample is weighed into a clean, labeled, preweighed 10 ml beaker. The beaker is placed in a forced air oven at approximately 75 degrees Celsius for 24 hours. The beaker with the dry sample is then weighed and the % dry weight is calculated by the formula:

(wt. dry sample and beaker) - (wt. beaker) (100)

Page: 61

#### **ANALYTICAL METHODS (Cont.)**

Method
Code Method Description
OO3

(wt. wet sample and beaker) - (wt. beaker)

007 LABORATORY: Geochemical & Environmental Research Group, Texas A&M

#### Trace Metal-Sediment

VII. Sediments are digested with aqua regia (3:1 HCI:HNO3) in glass beakers on a hotplate and diluted to volume with distilled water. Metals in the digestate are determined by 3 techniques, depending upon concentration and element. Mercury is determined by cold vapor atomic absorption spectrometry (AAS), in which Sn2+ is used to reduce HgO. Arsenic, selenium, cadmium, and lead are determined by graphite furnace AAS, in which electrical heating is used to produce an atomic cloud. Remaining elements (and Cd or Pb when in high concentration) are determined by atomic emission using an argon plasma.

013 LABORATORY: Geochemical & Environmental Research Group, Texas A&M

Acid Volume Sulfide (AVS)

XIII. Acid volatile sulfide (AVS) is defined as sulfides that are converted to H2S upon exposure to 1N HCl at room temperature for 1 hour. Approximately 10 grams of wet sediment are acidified with 1N HCl, and reactive sulfides that are converted to gaseous H2S are trapped as Ag2S and measured gravimetrically.